

REVIEW

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Diversification and domain evolution of molluskan metallothioneins: a mini review

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Abstract

Background: Metallothionein (MT) is a multifunctional protein playing important roles in homeostatic regulation and detoxification of metals. Mollusk species have been considered as useful sentinel platforms for MT-based biomarker approaches, and they have been reported to display an extraordinary structural diversity of MT proteins. However, potential diversity of molluskan MTs has not been fully explored and recent updates have suggested the need of revision of evolutionary hypothesis for molluskan MTs.

Results: Based on bioinformatic analysis and phylogenetic evidences, novel divergence mechanisms and paths were hypothesized in both gastropod and bivalve MT groups. Our analyses are suggestive of the taxon- or lineage-specific domain multiplication/duplication from the ancestral or prototypic MT. Diversification and selection of molluskan MTs might be driven by the needs for acquiring metal selectiveness, specialized novel function, and improved capacity of metal detoxification under environmentally stressed conditions.

Conclusion: The structural diversity and variations of molluskan MTs are significantly larger than previously understood. Undoubtedly, molluskan MTs have undergone dynamic divergent processes in their evolutionary histories, giving rise to the great diversity of domain structures in extant MT isoforms. Novel evolutionary paths for molluskan MTs newly proposed in this review could shed additional light onto the revision of the hypothesis for evolutionary differentiation of MTs in the molluskan lineage.

Keywords: Metallothionein, Phylum Mollusca, Structural diversity, Domain evolution

Background

Metallothionein (MT) is a metal-binding protein identified in almost living organisms. This housekeeping protein is a multifunctional, non-enzymatic effector playing important roles in various criteria of physiology of organisms under basal and stressed conditions (Carpenè et al. 2007; Mao et al. 2012). Owing to the high inducibility of its expression upon metal exposure, MT has been long given attention as one of core suits for biomarkers to address risks associated with metal-related environmental problems (Sarkar et al. 2006). Besides its fundamental roles in the homeostatic regulation of essential metals and detoxification of trace metals, MT plays important roles in host protective pathways under environmentally or physiologically perturbed conditions (Inoue et al. 2009; Chiaverini and Ley 2010; Lynes et al. 2014).

Benthic mollusks have been proposed as the useful sentinel platforms for biomarker approaches to aquatic and marine environments, in relation with their high bioaccumulation capacity of chemical elements from both water and sediment (Amiard et al. 2006; Geffard et al. 2007; Le et al. 2016). Further, their sedentary nature also makes it possible that biomagnification effects of the pollution could be effectively visualized without a significant consideration of complex migratory factors in the interpretation of bioaccumulation data (Gupta and Singh 2011). Accordingly, the exploitation of genetic determinants of molluskan MTs has been a progressively growing domain in the field of MT researches. To date, a number of previous literatures have claimed that molluskan species should represent a great structural diversity of MT proteins (Jenny et al. 2004; Jenny et al. 2006; Leignel and Lulier 2006). Moreover, some mollusk species particularly including American oyster *Crassostrea virginica* have shown extraordinarily large-sized MT isoforms comprising of over 100 amino acid (aa) residues (Jenny et al. 2004;

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Jenny et al. 2006; Tanguy and Moraga 2001), which have not been usually observable in vertebrate orthologs (Blindauer and Leszczyszyn 2010; Serén et al. 2014). Based on this, it has been widely proposed that the mollusks might have undergone evolutionary unique history in the divergence of MT proteins (Serén et al. 2014; Isani and Carpenè 2014; Wang et al. 2014; Jenny et al. 2016). Undoubtedly, structural diversifications would confer functional variations on these molluskan MTs, leading species-specific adaptation to environmental changes. Even yet fully elucidated, the prevalence of diversified isoforms are also likely in relation with significant variabilities and inconsistencies in the responses of molluskan MTs to metal or other stress exposures (in both laboratories and fields) (Amiard et al. 2006; Le et al. 2016).

Evolution of molluskan MTs has been proposed to be fundamentally based on the duplication events of their metal-binding domains. Domain duplication(s) from a common ancestral MT (a singular domain MT) might have given rise to multi-domain structured MTs (Jenny et al. 2004; Palacios et al. 2011). The prototypic MT has been supposed to go through further diversification into various distinguished isoforms in different taxa. This evolutionary theory has been comprehensively addressed with several well-known molluskan models such as oysters, mussels and air-breathing snails (Leignel and Laulier 2006; Jenny et al. 2016; Palacios et al. 2011; Aceto et al. 2011; Leung et al. 2014). However, in contrast to rich information on these popularly studied species, divergent processes of MTs in other mollusk species (in both gastropods and bivalves) have been quite limitedly explored yet. As the research on molluskan MTs progresses, it is becoming increasingly evident that structural variations and deviations of MT domains in this phylum may be significantly larger than previously understood. Recently, there have been considerable efforts on the isolation of novel MTs or MT-like proteins from different molluskan taxa. Now it is not difficult to find certain molluskan MT isoforms of which domain structures are hardly assigned into one of traditionally described categories. Hence, with this viewpoint, we aimed to review the structural diversity of molluskan MT domains with an angle to shed additional light onto the evolutionary path of MT domains in the molluskan lineage.

Status of MT isoform sequence data in public database

Currently the number of publically released MT gene sequences (both genomic and mRNA sequences; <https://www.ncbi.nlm.nih.gov/genbank/>) from the mollusk species has exceeded over 100 excluding the partial sequences or untrimmed ESTs. Almost all molluskan MT genes have been obtained from bivalves and gastropods, whereas no characterized MT sequence has been

reported yet from classes such as Aplacophora, Monoplacophora, Polyplacophora and Scaphopoda. No cloned MT sequence is available in the class Cephalopoda, barring couples of uncharacterized sequences predicted from the octopus (*Octopus bimaculoides*) genome scaffold. Hence, it is evident that genetic determinants of MT have been yet narrowly explored in the phylum Mollusca.

Many gastropod and bivalve species possess multiple paralogue MT isoforms. American oyster *Crassostrea virginica* (Ostreidae; Bivalvia) is the top species showing the greatest number of coexisting MT sequences in a single species ($n = 17$; non-redundant paralogue sequences at aa levels) (Jenny et al. 2004; Jenny et al. 2016); this isoform number is also the highest among all metazoans. Currently, in the Bivalvia class, 34 species belonging to four subclasses (Pteriomorphia, Anomalodesmata, Heteroconchia, and Palaeoheterodonta) have been recorded to reveal MT isoform(s) under GenBank accession codes. However, the species distribution for GenBank-deposited MT sequences in bivalves is highly skewed, in which only a few popularly studied species have dominated MT sequences. More than 50% of the total MT sequences have been from only the top five species belonging to either Mytilidae (mussels) or Ostreidae (oysters). Meanwhile, in the Gastropoda class (the largest and primitive class in the phylum Mollusca), only less than 20 non-redundant full-length MT sequences have been exploited from twelve different species belonging to one of seven families. These gastropod species include air-breathing pulmonate snails belonging to the Heterobranchia (species from four families, Planorbidae, Physidae, Helicidae, and Bradybaenidae), abalones (Haliotidae; Vetigastropoda), a limpet (Fissurellidae; Vetigastropoda), and a periwinkle (Littorinidae; Caenogastropoda). The species information and the accession code for each MT sequence used in this review are referred to in Additional file 1: Table S1.

A number of previous publications have reportedly indicated that most vertebrate MTs including mammalian and teleostean MTs may exhibit well-conserved primary structures. They are typically characterized by common features in polypeptide length (60~65 aa), calculated molecular weight (6~7 kDa), proportion of cysteine (Cys) residues (30~35%; arranged typically as Cys-X-Cys, Cys-X-X-Cys and Cys-Cys where the X is any non-Cys aa), and theoretical pI values (8.0~8.5) (Wang et al. 2014; Capasso et al. 2003; Cho et al. 2005; Cho et al. 2008; Cho et al. 2009) (Additional file 1: Figure S1A). Such a structural homology among vertebrate MTs may also attribute to a considerable degree of functional orthology between vertebrate species. However, it may not apply to the mollusk species. Remarkably variable or deviated scores in the primary structure have been frequently observable in molluskan MTs (Jenny et al. 2004; Tanguy and Moraga

2001; Baršytė et al. 1999; Tanguy et al. 2001) (Additional file 1: Figure S1B). First, molluskan MTs reveal often significant variations in polypeptide lengths among paralogue and orthologue isoforms. Median and mean values for the number of aa residues estimated from all the mollusk MT polypeptide sequences addressed in this study ($n = 109$) are 73.0 (7.2 kDa) and 80.2 (7.8 kDa) aa, respectively. Overall, these values are higher than that estimated with representative vertebrate MT orthologs. More noticeably in molluskan group, considerable numbers of MTs represent extraordinarily short or long lengths apparently deviated from the mean (and median) value above. Within the Bivalvia class, the length of MT polypeptides ranges from 43 aa (4.2 kDa) to 204 aa (20.3 kDa); noticeably both are recorded as paralogs from a single species *C. virginica* (Ostreidae) (GenBank accession numbers, AY331700/AY331701 and AY331706, respectively). Other oyster species also display the significant variations in the sizes of MT isoforms. However, the mussel species belonging to the order Mytiloida represents relatively a uniform feature in the range of the MT protein lengths (66~75 aa). Besides the two main bivalve taxa, it is worthy to note unusually long MT isoforms reported from *Argopecten irradians* (Pectinidae; EF093795, and EU734181) and *Pisidium coreanum* (Sphaeriidae; GQ268325).

On the other hand, mean and median values for MT polypeptide lengths available for gastropod MTs were 67.5 and 65.0 aa, respectively. Currently, the shortest gastropod MT sequence is found to be *Physa acuta* (Physidae; GU259686) MT consisting of 59 aa, whereas the longest one is the MT isoform with 100 aa reported in a periwinkle species *Littorina littorea* (Littorinidae; AY034179). Except these shortest and longest sequences, the polypeptide lengths of other gastropod MTs are found to fall in the range from 64 to 70 aa. Similarly with bivalves, multiple isoform MTs ($n = 2\sim 4$) have been reported in pulmonate snails belonging to Helicidae or Planorbidae, while marine gastropods including abalones have shown only a single MT isoform; yet still unclear if they have additional MT isoform gene(s) or not (Additional file 1: Table S1; Additional file 1: Figure S1).

Second, molluskan group shows a wider range of distribution for theoretical isoelectric point (pI) values across MT isoforms as compared to mammalian and teleostean MT groups (Additional file 1: Figure S1). In mammals, MT-IIIs are only the isoform family (but excluding murine MT-IIIs) to show a significantly acidic pI values (4.79~4.82), while most other isoforms belonging to MT-I, -II or -IV class typically represent pI values higher than at least 7.5. Most teleostean MTs show pI values ranging from 7.8 to 8.4. On the other hand, bivalve MTs display pI values ranging 4.31 to 8.56 depending on isoforms. In the Gastropoda group, the pI values of the MT proteins range from 5.16 to 8.28. Although

the numbers of taxa available for MTs from Gastropoda class were currently limited, theoretical pI values estimated from the category group Vetigastropoda (pI 5.16~6.00) were relatively lower than those from Heterobranchia (pI 6.10~8.28). Possibly, MT proteins with largely different pI values might exhibit differential capability to bind or interact with charged molecules in the host protective process. In addition, contents of some positively charged aa (e.g., Lys) may influence importantly the metal-binding function of MT protein. These aa residues (particularly at positions in vicinity to Cys) are thought to be involved in the stabilization of the interaction between MTs and metal ions through the electrostatic interactions to bridge the protonated basic residues and the negatively charged metal-thiolate complex (Pedersen et al. 1994). Thereby, such a wide range of pI values may be potentially suggestive of, at least in part, dynamic diversification and subfunctionalization among molluskan MT isoforms, although the biological and/or functional implications of such a wide range of pI values should be explored in the future.

Third, molluskan taxa are found to have a tendency of relatively lower Cys content (mean \pm s.d. = $28.1 \pm 1.8\%$) in their MT proteins than are mammalian ($32.4 \pm 1.3\%$) or teleostean ($33.3 \pm 0.3\%$) groups. Also, the intra- and interspecies variations in Cys contents of MTs are larger in mollusks (especially in bivalves; ranging 21.2~31.9%) than in mammals (26.2~34.4%; 31.1~34.4% if MT-IIIs excluded) and fish (32.8~35.0%) (Additional file 1: Figure S1). Metal binding property and capacity of molluskan MTs have been studied in only a few species (Palacios et al. 2011). However, Cys residues are known to be essential for the affinity to bind metal ions where the number of metal ions bound by the MT may be fundamentally determined by the number of Cys residues (Amiard et al. 2006; Jenny et al. 2016; Vergani et al. 2005). Hence, it is able to suggest that molluskan MTs may display relatively larger variations in the metal binding capacity among isoforms than vertebrate orthologue groups.

Taken together, all the three parameters above mentioned are undoubtedly indicative of large structural variations and divergence of MT proteins among molluskan species. Based on this overview, taxa (or lineage)-specific patterns for structural diversification of MTs are described more details in following sections, with a particular attention on the arrangement of Cys motifs.

Structural diversity of MT isoforms in major molluskan taxa

Nomenclature and classification of MTs used in this paper were referred to GenBank (NCBI) based on the definition of each sequence. If the MT sequence was published in scientific paper(s), its classification was checked again. Within a given species, the redundant

MT sequences at aa level were not included in analyses (Additional file 1: Table S1). Classification of putative domain structure in each MT sequence was based on the number and arrangement pattern of Cys motifs as described previously (Jenny et al. 2016), since there have been no empirical studies on three-dimensional structures of molluskan MT proteins. In general, domain structure in MT is designated α and β (Braun et al. 1986; Binz and Kagi 1999). Usually, the α -domain contains eleven to twelve Cys residues, binds four divalent metal ions and confers structural stability on the MT polypeptides. On the other hand, the β -domain, binds three divalent metal cations through the nine Cys residues and participates in metal exchange reactions via glutathione-shuttling with metal-requiring apoproteins (Jiang et al. 1998; Jiang et al. 2000).

Gastropoda MTs

In a total, nineteen non-redundant MTs (from twelve species belonging to seven families) including a putative MT predicted in the unplaced genomic scaffold sequence (*Biomphalaria glabrata*; Planorbidae) were analyzed with sequence alignments. Among the nineteen sequences, 17 sequences with 59 to 70 aa residues are found to be fairly aligned in the multiple sequence alignment trials. In spite of substantial differences in non-Cys residues among taxa, they share a conserved pattern of Cys motifs. Eighteen Cys in these gastropoda MTs are arranged as [Cys-X-X-Cys \rightarrow (Cys-X-Cys)₃ \rightarrow Cys] \rightarrow [(Cys-X-Cys)₂ \rightarrow Cys \rightarrow (Cys-X-Cys)₂] (Additional file 1: Figure S2). It indicates that gastropoda MTs represent the protein structure comprising of two distinguished β -domain forms (i.e., $\beta_2\beta_1$ -form) designated by the recent suggestion to propose the presence of two hypothetical ancestral β -domains (Jenny et al. 2016). The β_2 -domain structure at the N-terminal of gastropod MT protein is similarly observed in *C. virginica* MT-IIIs as well as in the β -domains of vertebrate (mammals and fish) MTs. On the other hand, the C-terminal β_1 -domain of the gastropod MT is commonly found in various molluskan MTs. According to the $\beta_2\beta_1$ -structural scheme, the shortest *P. acuta* MT (comprised by 59 aa) is thought to have lost two Cys in its β_2 -domain. In that alignment, MTs from Vetigastropoda species possess more aa residues (5 or 8 aa) in the intervening region between the β_2 - and β_1 -domains than those from Heterobranchia species (2 aa).

Besides the common $\beta_2\beta_1$ -shape, gastropod group represents two significantly lengthy MT polypeptides. One is 100-aa MT from *Littorina littorea* (Caenogastropoda; AY034179) (English and Storey 2003) and the other is 124-aa MT (XP_013080485; deduced from the unplaced genomic scaffold) from *B. glabrata* (Additional file 1: Figure S2). Based on manual alignment, *L. littorea*

MT is proven to show a conserved pattern for Cys arrangements in N- and C-terminal parts (i.e., nine Cys respectively in putative N- and C-terminal domain regions). Considering the Cys motif patterns, the N- and C-terminal parts of the *L. littorea* MT could be designated β_2 - and β_1 -domains, respectively. Further, a closer examination on the intervening region (comprised by 32 aa) between the β_2 - (N-terminal) and β_1 - (C-terminal) domains has indicated that the 32-aa internal segment has been potentially a duplicated copy of the N-terminal β_2 -domain. It conserves clearly 9 Cys residues and shows a considerably high sequence identity (75%) to the N-terminal (β_2 -domain) domain. Based on our peer review, the structure of *L. littorea* MT could be considered as a novel shape of gastropod MT characterized by $\beta_2\beta_2\beta_1$ -domain form. Hence, this newly proposed structure suggests that domain duplication event might have served as a driving force to figure the large MT in certain gastropod taxa.

Another example for the domain duplication in gastropod MT is the 124-aa *B. glabrata* MT (XP_013080485). In that MT polypeptide sequence, three putative domain regions sharing a considerable sequence similarity in one another could be identified, and each of the three putative domains may be designated β_2 -structure based on their Cys arrangement patterns (Additional file 1: Figure S2). As numbered from the N-terminal, the first β_2 -domain and the second β_2 -domain share the conserved Cys motif frame (except one additional Cys in the second β -domain). They also show the high sequence homology (76.5%) each other. The putative third β_2 -domain linked to the second β_2 -domain is found to display 66.7% of sequence homology to the first and second β_2 -domains. Within this context, the *B. glabrata* MT could be proposed to possess at least three duplicated β_2 -domains tandemly arrayed in a tail-to-head fashion. On the other hand, the remaining C-terminal part (23-aa) following the $\beta_2\beta_2\beta_2$ -domain region is found to contain five Cys (three singlet Cys and a Cys-Cys doublet motif). Unlike the *L. littorea* MT above, the C-terminal region of this *B. glabrata* MT display no typical shape to be categorized into one of known domain structures (Additional file 1: Figure S2). Currently, the origin of this C-terminal region has been unknown. Further validation of scaffold genomic sequences along with mining of similarly organized MTs from other Heterobranchia genomes would be needed to get a deeper insight into the mechanism responsible for the formation of the array of three β_2 -domain region in this gastropod MT. This pulmonate snail species has already been known to express functionally diversified (i.e., different metal selectiveness) MT isoforms [(i.e., Cd-MT (GQ205374), Cu-MT (GQ205373) and intermediate Cd/Cu-MT (GQ205375)] (Berger et al. 1997). Hence, it would also be valuable to examine the expression patterns of this newly identified MT regarding its potential differentiation in physiological

function and/or metal responsiveness in comparison with previously characterized paralogs from this species.

Bivalve-ostreidae MTs

Structural diversity of MT families in Ostreidae has been comprehensively described with *C. virginica* model (Jenny et al. 2004; Jenny et al. 2006). Currently, seventeen *C. virginica* MT sequences available in GenBank could be classified into one of four MT isoform families (MT-I, -II, -III, and IV). The MT families consist of 2 (MT-IA and MT-IB), 8 (MT-IIA, MT-IIB, MT-IIC, MT-IID, MT-IIE, MT-IIF, MT-IIG, and MT-IIH), 3 (MT-IIIA, MT-IIIB, and MT-IIIC), and 3 (MT-IVA, MT-IVB, and MT-IVC) subisoforms, respectively. In addition to these 16 sequences, one MT sequence named MTA (GenBank accession no. AF506977) is independently recorded in this oyster species (see Additional file 1: Table S1). Of the 16 isoforms, the prototypical MT structure (corresponding to MT-I form; MT-IA and MT-IB) is 75-aa MT possessing 21 Cys residues (28% of Cys content). The MTA isoform can also be classified to the same prototype (i.e., MT-I) with an addition of Asp in the region between 16th and 17th Cys residues. They share a high sequence homology one another in a prototypic $\alpha\beta_1$ -domain-structure (Jenny et al. 2004). This $\alpha\beta_1$ -structure is well conserved also in other oyster species, including *C. gigas* (Pacific oyster; AJ242657), *C. ariakensis* (the Suminoe or Asian oyster; DQ342281) and *C. rivularis* (the Jinjiang oyster; JN225502). The same domain structure is also relevant with isoforms from non-*Crassostrea* species *Ostrea edulis* (Tanguy et al. 2003). However, relative large substitutions of non-Cys aa (also the replacement of three Cys with other aa in the *O. edulis* MTb isoform) have been found in the *O. edulis* MTs compared to MT-I orthologs from *Crassostrea* species (Additional file 1: Figure S3A).

C. virginica MT-II family includes eight subisoforms (MT-IIA to MT-IIH), and this MT-II family has been known to be classified into two subgroups. First, the MT-IIA/-IIB group possesses a sole α -domain of which sequence is highly conserved with that of the prototypic $\alpha\beta_1$ -MT-I. The loss of functional β_1 -domain has been proposed to occur due to the point mutation in the linker region (i.e., non-sense mutation resulting in a stop codon). Consequently, MTs belonging to this group reveal noticeably short polypeptide length (43 aa) (Jenny et al. 2004). Second, on the contrary, the remaining six *C. virginica* MT-II isoforms are lengthy polypeptides (94-aa MT-IIC, 149-aa MT-IID/-IIE, 145-aa MT-IIF, 204-aa MT-IIG, and 200-aa MT-IIH). They have tandemly duplicated copies of α -domain where the numbers of repetitive duplications are variable among isoforms. In MT-IIC, only one duplication event is predicted (i.e., two tandem duplicate copies), while subisoforms MT-

IID/-IIE/-IIF and MT-IIG/-IIH display tandem arrays comprised of three and four α -domain copies, respectively (Jenny et al. 2004). Between and among duplicated (repetitive) copies, a few aa substitutions have been found. Collectively, the divergent process of *C. virginica* MT-II family has occurred through the loss of β_1 -domain (e.g., MT-IIA/-IIB; α -domain-structure) followed by further divergence into various subisoforms having differential numbers of duplicated α -domains (e.g., MT-IIC to MT-IIH; $\alpha_{(n=2\sim4)}$ -domain structure) (Jenny et al. 2016).

However, this divergence pattern has not been always a common finding in the Ostreidae lineage (Additional file 1: Figure S3A). Rather than the loss of β -domain, *C. gigas* MT-II family has represented the tandem duplication of 32-aa β_1 -domain with retaining the α -domain, giving rise to the $\alpha\beta_1\beta_1$ -domain structure (Tanguy and Moraga 2001). Further, unlike in *C. virginica*, there has been no variation in repeat numbers among *C. gigas* MT-II subisoforms. MT (AF349907) from another *Crassostrea* species, Portuguese oyster (*C. angulata*) has exhibited the duplication of only a short, partial β_1 -domain fragment (7-aa region), giving rise to a non-canonical $\alpha\beta_1\beta_{1P}$ -structure with truncated C-terminus. Because there has been no other publically released MT paralog from *C. angulata*, it has been yet unclear if this oyster species may possess any paralog copies representing the complete $\alpha\beta_1\beta_1$ -domain structure or not. Beyond the *Crassostrea* genus, our survey against GenBank has identified that *Alectryonella plicatula* MT (KP875559; 107-aa) should also display the typical $\alpha\beta_1\beta_1$ -domain structure. Moreover, the *A. plicatula* MT shows very high sequence homology to its *Crassostrea* orthologs (MT-IIIs), indicating the common origin of this multi-domain structure. Currently, the known $\alpha\beta_1\beta_1$ -structured MT subisoforms in Ostreidae (except for the *C. angulata* MT with a truncated C-terminus) share the same N-terminal (Met-Ser-Asp-Pro) and C-terminal (Cys-Lys-Lys) motif residues (Additional file 1: Figure S3A).

On the other hand, the *C. virginica* MT-III group consists of three homogenous subisoforms MT-IIIA/-IIIB/-IIIC. They share each other high sequence identity including 18 Cys residues, and the distribution pattern of the 18 Cys have been proposed as the array of two β -domains as $[(\text{Cys-X-Cys})_4 \rightarrow \text{Cys}] \times 2$ (i.e., $\beta_2\beta_2$ -MT) (Additional file 1: Figure S3B). The arrangement pattern of nine Cys is obviously similar with the β_2 -domain of the gastropod $\beta_1\beta_2$ -MTs (Jenny et al. 2006; Jenny et al. 2016). The same $\beta_2\beta_2$ -domain structure has also been found in *C. gigas* MT-III (JF781299); however unlikely in *C. virginica*, multiple MT-III subisoforms have not been characterized in *C. gigas* (Cong et al. 2012). The *C. gigas* MT-III shows a series of substitutions of non-Cys aa from the *C. virginica* MT-III isoforms. The overall

sequence identity of MT-IIIs between the two *Crassostrea* species is about 70% (Additional file 1: Figure S3B). Meanwhile, *Crassostrea* MT-III isoforms reveal considerably low pI values (4.38~4.77 for *C. virginica* and 4.31 for *C. gigas*) as compared to other MT isoforms showing pI values > 7.5. With the viewpoint of the low pI value, *C. virginica* MT-III may resemble mammalian MT-III family. Most known mammalian MT-III isoforms except murine MT-IIIs reveal acidic pI ranges (4.79~4.82) with acidic 6-amino-acid insert in the C-terminal region. Synthesis of mammalian MT-III is not inducible by heavy metals and localized predominantly in the central nervous system (Faller 2010). Unique roles of mammalian MT-III differing from other MT isoforms have been characterized as the neuronal growth-inhibitory factor to inhibit neuronal outgrowth (Wang et al. 2006). Specific roles of bivalve MT-III differing other MT family groups have not been yet extensively addressed. However, the expression study with *C. virginica* MT-III has indicated that the *C. virginica* MT-III showed quite a low basal level of expression in adult tissues (i.e., only actively expressed in early larvae). Further, *C. virginica* MT-III represented only a moderate responsiveness to heavy metal exposures in both larvae and adults (Jenny et al. 2006). However, on the contrary, the *C. gigas* MT-III has been reported to be significantly induced by zinc (as a main regulator for zinc homeostasis), and it may be a participating member for cadmium detoxification in the adult tissues (Cong et al. 2012). Thereby, it suggests the functional differentiation/divergence of MT-III isoforms during speciation events in the *Crassostrea* lineage. Within the context of this hypothesis, it is worthy to have an attention on another *C. gigas* MT isoform named mt3 under the accession number AJ295157. This *C. gigas* mt3 isoform is not a true MT-III family member with the $\beta_2\beta_2$ -domain structure. Rather than, mt3 should be considered as a MT-I member because it represents the $\alpha\beta_1$ -domain structure (see Additional file 1: Figure S3A). However, due to the significant non-Cys aa substitutions, *C. gigas* mt3 displays only 67% sequence identity to its paralogue MT-I isoform. Moreover, the substitutions include the change of three Lys residues to uncharged aa as well as the replacement of uncharged aa with negatively charged aa. Such aa substitutions might give rise to lower pI value (5.98) of mt3 than those of its paralogue MT-I members. A previous study has reported that mt3 should have the extremely low basal expression level with only moderate or minute responsiveness to metal exposure (Marie et al. 2006). The mt3 has been suggested to have probably no significant physiological functions under metal exposure and to be expressed only in particular developmental stages (Marie et al. 2006). Hence, taken together, it could be hypothesized that *C. gigas* mt3 might have been

diverged from the prototypic MT-I through the non-Cys aa changes. This isoform is likely to show, at least in part, certain functional orthology to the *C. virginica* MT-III, although they have different domain structures.

Finally, the MT-IV isoforms (MT-IVA, MT-IVB and MT-IVC; 83 aa in length) from *C. virginica* have been proposed as variant forms of $\alpha\beta_1$ -MT. This isoform group has been supposed to have experienced a series of aa substitutions including Cys residues, giving rise to 25 Cys with the formation of a Cys-Cys doublet (in the α -domain) and three Cys-Cys-Cys triplet motifs (in the β_1 -domain). The C-terminal residue of the *C. virginica* MT-IV isoform (glutamine-alanine-threonine) is also notably different from those of other paralog isoforms. Thereby, the proposed designation of domain structure for MT-IVs could be $\alpha'\beta_1'$ -form. In addition to *C. virginica*, two *Crassostrea* species (*C. gigas* and *C. ariakensis*) possess MT-IVs of which primary domain structures are fairly conserved with that of *C. virginica* MT-IV. However, *C. gigas* MT-IV (87 aa; AM265551) and *C. ariakensis* MT-IV (86 aa; JF919323) represents one additional Cys residue at C-terminal region (Additional file 1: Figure S3B). Besides the above *C. gigas* MT-IV clone (AM265551), genome sequencing of *C. gigas* (Zhang et al. 2012) represents two unplaced genomic scaffolds [scaffold852 (JH816574) and scaffold1297 (JH818394)] each containing the putative gene encoding MT (EKC32371 and EKC28510, respectively, in the two scaffolds). The deduced sequences of these MTs are 128 and 137 aa in length, respectively. Both MTs are predicted to possess unusual N-terminal regions (41-aa for EKC32371 and 50-aa for EKC28510). However, immediately following the N-terminal region, the two putative MTs represent the 87-aa-structure that is apparently homologous to the *C. gigas* MT-IV isoform (AM265551). When the unusual N-terminal regions are excluded from these two MT-IV-like sequences, the three MT sequences (one characterized MT-IV and two in-scaffold sequences) reveal only one aa substitution, although the scaffold sequences should be further validated in future.

Bivalve-mytilidae MTs

In Mytilidae, 33 full-length, non-redundant MT aa sequences were retrieved from five taxa (12 sequences from *Mytilus edulis*, 6 from *M. galloprovincialis*, 2 from *Mytilus* sp., 11 from *Perna viridis* and 2 from *Bathymodiolus azoricus*). Mytilid MTs are a structurally more homogeneous group as compared to *Ostrea* MT group. They represent 66-aa to 75-aa polypeptides containing 19 to 23 Cys residues, and display typically the $\alpha\beta_1$ -domain structure (Additional file 1: Figure S4).

In the mytilid mussels, two types of MT isoforms, MT10 and MT20 have been described previously (Aceto et al. 2011; Leung et al. 2014). These two MT types

differ in mass and Cys arrangement. The monomeric form MT10 (~10 kDa) represents generally 73-aa polypeptides including 21 Cys residues mainly arranged as nine Cys-X-Cys motifs. On the other hand, the dimeric form MT20 (~20 kDa) is typically 72-aa polypeptides containing 23 Cys residues. Unlike MT10, the MT20 isoforms show a Cys-Cys doublet in the α -domain. MT10 and MT20 have been reported to be functionally differentiated. MT10 has been found to be more abundant than MT20, and hence it could be considered as a main player for the regulation of homeostasis under basal conditions (Leung et al. 2014; Lemoine et al. 2000). Under metal-exposed conditions, MT10 and MT20 have been known to display differential responses and binding ability to essential and non-essential metals. MT10 could be actively inducible by various metals while MT20 be more preferentially associated with non-essential metals such as Cd and/or Hg (Raspor et al. 2004; Dondero et al. 2005; Vergani et al. 2007).

From multiple sequence alignment, mytilid MTs could be categorized into three main groups (Additional file 1: Figure S4). The first group consists of three MT10B sequences (two from *M. edulis* and one from *M. galloprovincialis*; AJ577126, AJ577127 and DQ848984, respectively). They represent 66-aa polypeptides containing 19 Cys residues with the deletion of one Cys-X-Cys motif in the α -domain. All the three MT sequences are originated from intronless MT genes (Leignel et al. 2005; Yang et al. 2014). The second group contains twenty-four MT10 isoform sequences. They reveal 72 to 75 aa in lengths and possess 21 Cys residues in conserved positions (Lemoine et al. 2000; Khoo and Patel 1999). There are only two exceptions; one is the replacement of a Cys with Arg in *P. viridis* MT-IA (JN596471) and the other is an insertion of an additional Cys in *P. viridis* MT (AF036904). Within the second group, the MTs are likely to be sub-grouped according to known taxonomic appraisal at genus level (i.e., *Bathymodiolus*, *Mytilus* and *Perna*). All the *P. viridis* MT-II isoforms (named MT-IIA, -IIB, -IIC and -IID) are found to possess 72 aa, as similarly with the mytilid MT20 isoforms. Nevertheless, based on their Cys motifs frame, these *P. viridis* MT-IIs (JN596477 to JN596480) have been proposed as MT10 members. Previous phylogenetic analysis has claimed an early divergence of *P. viridis* MTs from the main mytilid MT10/MT20 groups (Leung et al. 2014). When aligned with other mytilid orthologs, *P. viridis* MT10-I and/or MT10-II represent several residues distinct from other mytilid MT isoforms. They include positions 11th (alignment position; Gln/Lys in *P. viridis* MTs vs. Asn in all other mytilid MT10 and MT20 isoforms), 62nd (Gln vs. Gly/Asp) and 74th (Ser vs. Gly). Further, *P. viridis* MT10 isoforms (both MT10-I and MT10-II) are found to share the same aa in several positions with mytilid MT20 isoforms. These could be exemplified by positions 34th

(alignment position; Ser in *P. viridis* MTs and MT20s vs. Gly in other mytilid MT10s), 39th (Gly vs. Lys), and 73rd (Ser vs. Pro). Finally, the third group is comprised of six MT sequences from three *Mytilus* species. They are 72-aa polypeptide containing 23 Cys residues (i.e., MT20 type). Only one exception is the substitution of Cys to Arg in the *M. edulis* MT-20 clone. From the alignment, two MT20-specific residues could be found at positions 24th (Lys in MT20s vs. Glu in all MT10s) and 68th (Asn vs. Thr) (Additional file 1: Figure S4). In particular, the change from negatively charged aa (e.g., Glu) to Lys at position 24th is likely related with the MT20-specific formation of Cys-Cys doublet motif, since the positively charged aa (e.g., Lys) at vicinity to Cys motif is considered to play important roles in the stabilization of the metal binding reaction in most MT proteins (Pedersen et al. 1994).

Bivalve-other taxa

Besides the two main bivalve taxa (Ostreidae and Mytilidae), 27 non-redundant, full-length MT isoforms have been exploited from 21 bivalve species belonging to one of six orders Pterioidea, Arcoidea and Pectinoidea (subclass Pteriomorpha), Pholadomyoidea (subclass Anomalodesmata), Veneroidea (subclass Heteroconchia), and Unionoidea (belonging to Palaeoheterodonta). From the multiple sequence alignment, most of them are found to represent $\alpha\beta_1$ -domain-structure with a conserved 21-Cys-frame (Additional file 1: Figure S5A). However, several variant isoforms are also found to show modification(s) of Cys motifs in one or two positions, giving rise to the generation of Cys-Cys doublet, substitution, insertion or deletion. Several variant isoforms are found to retain the total number of Cys residues (i.e., 21 Cys) while others show changes of the total number of Cys residues. MT isoforms from *Pinctada maxima* (pearl oyster; FJ389580) (Tang et al. 2009) and *Laternula elliptica* (Atlantic clam; DQ832722/DQ832723) display the insertion of an additional Cys in the third Cys-X-Cys motif, resulting in the Cys-Cys-Cys motif at that position. On the contrary, an MT isoform from *Hyriopsis schlegelii* (freshwater pearl mussel; MT2; KJ019821) has lost one Cys residue at the second Cys-X-Cys motif along with considerable alterations in non-Cys aa residues. Unlike its paralog (*H. schlegelii* MT1; KJ019820), the *H. schlegelii* MT2 has been proposed as a genetically separated isoform (Wang et al. 2016). A recent study has indicated that these two *H. schlegelii* MT paralogs might have been subfunctionalized as evidenced by clearly distinct tissue expression patterns (i.e., constitutive expression of *H. schlegelii* MT1 vs. gonad-specific or predominant expression of *H. schlegelii* MT2) (Wang et al. 2016). Another example for the large difference between paralog MT isoforms is *P. martensi* MT isoforms. Even though the *P. martensi* MT1 (KC197172.1) represents the common $\alpha\beta_1$ -shape,

its paralog MT2 (KC832833.1) exhibits an apparently non-canonical pattern of Cys arrangement (20 Cys).

On the other hand, two MT isoforms from Veneroidea are found to have noticeably less number of Cys residues than others: one is the duck clam *Macra veneriformis* (Mactridae) MT with 18 Cys (59-aa; Cys content = 30.5%; FJ611963) (Fang et al. 2010; Fang et al. 2013) and the other is Venus clam *Cyclina sinensis* (Veneridae) MT with 16 Cys (74-aa; Cys content = 21.6%; HM246244) (Lü et al. 2012). The *M. veneriformis* MT seems to have lost an internal fragment near N-terminal region (possibly corresponding to the α -domain) containing three Cys residues (likely a Cys-X-Cys motif and one conserved Cys). The *C. sinensis* MT lacks a Cys-X-Cys motif in α -domain and additionally three Cys residues (Cys-X-Cys motif and one Cys residue) probably in the β -domain.

Importantly, three MT isoforms display large polypeptide sizes comprising of more than two putative domains (Additional file 1: Figure S5B). Of the three MTs, two MT sequences (MT1 and MT2) are from the bay scallop *Argopecten irradians* (Pectinidae) and remaining one isoform is from the fingernail clam *Pisidium coreanum* (Sphaeriidae). These sequences have been reported earlier but their domain structures have never been addressed clearly. Even though *A. irradians* MT1 (145-aa; 40 Cys; EF093795; (Liu et al. 2006)) and MT2 (110-aa; 28 Cys; EU734181; (Wang et al. 2009)) exhibit essential features of mollusk MTs (i.e., the presence of characteristic Cys-X-Cys motifs), their overall structures are more or less complicated and difficult to be simply categorized into one of currently known shapes of bivalve MTs. However, in a broad sense, these isoforms may bear a resemblance to the MT isoforms with multi- β -domain-structure. For both *A. irradians* MT isoforms, the C-terminal region may be considered the β_1 -domain possessing nine Cys as $(\text{Cys-X-Cys})_2 \rightarrow \text{Cys} \rightarrow (\text{Cys-X-Cys})_2$. In addition to the C-terminal β_1 -domain, *A. irradians* MT1 potentially exhibits three tandemly arrayed β_2 -like domains. However, each β_2 -like domain in the *A. irradians* MT1 displays some non-canonical arrangement of Cys. First, two Cys of the first Cys-X-Cys motif in each domain is separated further by intervening 2~4 aa residues (i.e., similar with the pattern found in N-terminal regions of gastropod β_2 -domains). Second, Cys-Cys doublet motifs rather than a canonical Cys-X-Cys motif are present in the first and third β_2 -like domains. Third, an additional Cys-X-Cys motif exists in the flanking regions between the first and second β_2 -like domains as well as between the second and third β_2 -like domains. Nevertheless, the overall shape of *A. irradians* MT1 may be designated $\beta_2\beta_2\beta_2\beta_1$ -like structure, although this novel proposal should be further challenged with empirical structural analyses. Sequence comparisons among/between these successive β_2 -like

domains indicate that they share little sequence similarity one another except conserved Cys residues (Additional file 1: Figure S5B). Within this scheme, the *A. irradians* MT2 could be treated as a paralog having the less number of β_2 -domains (i.e., putatively designated $\beta_2\beta_2\beta_1$ -like structure). The *A. irradians* MT2 also reveals some non-canonical attributes including the lack of one Cys residue and the formation of Cys-Cys doublet in the N-terminal β_2 -like domain. These two *A. irradians* MT isoforms are found to share only a little sequence homology, indicating that they may be quite distantly related paralogs (Wang et al. 2009).

On the other hand, the 105-aa *P. coreanum* (Sphaeriidae) MT (GQ268325; 31 Cys) (Baek et al. 2009) is found to show the domain multiplication to resemble the $\alpha\beta_1\beta_1$ -structure, of which Cys arrangement is similar with that of *C. gigas* MT-II. However, unlike *C. gigas* MT-II to show the tandem array of two homogenous β_1 -domains, *P. coreanum* MT contains the two heterogeneous β_1 -domains with no apparent sequence homology between the two domains. The *P. coreanum* MT lacks a common triplet linker sequence (Lys-Val-Lys/Val) between α - and first β_1 -domain. The array of two heterogeneous β_1 -domains linked to N-terminal α -domain observed in *P. coreanum* MT could be the novel structure of bivalve MT proteins (Additional file 1: Figure S5B).

Domain evolution in molluskan MTs

Current theory for domain evolution of molluskan MTs

Currently, the proposed hypothesis for the evolution of molluskan MT has been based on the domain duplication event(s) from an ancestral single domain-structured MT, in which the β -domain has been considered as the ancestral shape (Cols et al. 1999). After an early duplication event of the ancestral β -domain, the resultant $\beta\beta$ -domain MT has undergone divergent processes, given rise to the $\alpha\beta$ -structure in certain taxa (Jenny et al. 2016; Braun et al. 1986; Cols et al. 1999). Difference in the metal-binding properties between the α -domain and the β -domain makes the two domains to represent differentiated roles in the cellular physiology. Generally, α -domain plays a more prevalent role in Zn homeostasis and detoxifying sequestration of toxic metals (e.g., Cd) whereas the β -domain is primarily responsible for the homeostatic regulation of essential metals (e.g., Cu) (Jenny et al. 2004; Cols et al. 1999; Nielson and Winge 1984; Xiong et al. 1998). Consequently, the multi-domain MT in specific taxa acquiring both α - and β -domains was able to perform the dual functions; the detoxification of toxic metals by the α -domain and the homeostasis of physiologically relevant metals by the β -domain (Jenny et al. 2016; Cols et al. 1999; Nielson and Winge 1984; Nielson and Winge 1983).

The latest phylogenetic work has proposed that two distinct ancestral β -domains (designated β_1 and β_2 domains) might have existed and given rise to the structural diversity of all molluskan MTs (Jenny et al. 2016). In that literature, they have hypothesized separate paths of the evolution of the two ancestral MTs in the major taxa within the mollusk phylum. The two β -domains appear to have diverged into two structurally different MT isoform types (i.e., $\alpha\beta_1$ -MT and the $\beta_2\beta_2$ -MT) in bivalves whereas in gastropods, the two ancestral β -domains form a single structural $\beta_2\beta_1$ -MT isoform. With *C. virginica* model, the structural diversity of bivalve MT isoforms has been highlighted to demonstrate evolutionary paths from not only $\alpha\beta_1$ -domain but also $\beta_2\beta_2$ -domain (Jenny et al. 2016). On the contrary, in the gastropoda lineage, the $\beta_2\beta_1$ -domain structure has been proposed as a typical appearance common to most extant gastropod MTs. Instead of a series of domain duplications seen in bivalve MTs, gastropod MTs appears to have diverged to functionally differentiated isoforms (i.e., Cd-MT, Cu-MT or intermediate Cd/Cu-MT) through the composition changes of non-Cys aa residues (Jenny et al. 2016; Palacios et al. 2011; Cols et al. 1999). However, our bioinformatic analyses in this study suggest that the current theory on MT domain evolution in the phylum Mollusca could be revised based on newly recognized evidences. Novel hypothetical paths and additional insights into the domain evolution of molluskan MTs are proposed in following sections.

Novel evidences for domain duplication in gastropod MTs

The non-canonical domain structure of large MTs from two gastropod species (*L. littorea* and *B. glabrata*) may be considered as novel shapes. Phylogenetic analysis of gastropoda MT domains (β_1 or β_2) has generated the two major clades separated depending on the types of β -domains (β_1 or β_2) (Fig. 1). The gastropod β_2 -clade has been proven to contain all previously proposed gastropoda β_2 -domains together with the putative β_2 -domains of the large MTs proposed in this study. For the large *B. glabrata* MT, three β -domains (the first to third domains numbered from the N-terminal) are closely clustered together and placed in the major clade comprising the gastropod β_2 -domains. This result suggests obviously that they are duplicated copies of β_2 -domains that might have evolved through the tandem duplication events. On the other hand, the C-terminal region containing only five Cys is not clustered with any typically known β_1 or β_2 -domain sequences. Although we did not provide clear evidence for the origin of this C-terminal region, the most likely scenario is that the originally existed β_1 -domain at C-terminal in the ancestral $\beta_2\beta_1$ -MT might have undergone certain recombination(s) including the loss of some parts during duplication events of neighboring β_2

domains. Based on this assumption, the unusual C-terminal part of *B. glabrata* MT might be a reminiscent, partial segment (designated β_{1P} here) originated from the early β_1 -domain. Hence, the tandem duplications of β_2 -domain accompanied with partial loss of the C-terminal β_1 -domain may be a plausible mechanism to produce the current $\beta_2\beta_2\beta_2\beta_{1P}$ structure in this pulmonate species. We performed additional analyses on the duplicated β_2 -domains of this large MT (first and second β_2 -domains used for analysis) in order to hypothesize a plausible reason responsible for the happening of this evolutionary episode. For this, β_2 -domains of previously known MTs from pulmonate species (i.e., Cd-MT, Cu-MT and intermediate Cd/Cu-MT) were included in analyses together with the β_2 -domains of this large *B. glabrata* MT. From the sequence alignment, the duplicated β_2 -domains of the *B. glabrata* $\beta_2\beta_2\beta_2\beta_{1P}$ -MT revealed a more sequence similarity to Cd-MT than to Cu-MT and Cd/Cu-MT. Phylogenetic analysis of β_2 -domains from *B. glabrata* paralogs also showed a close relationship between the $\beta_2\beta_2\beta_2\beta_{1P}$ -MT and Cd-MT (Additional file 1: Figure S6). Although the tree topology on the affiliation was not statistically supported, it could be enough to hypothesize that the emergence of $\beta_2\beta_2\beta_2\beta_{1P}$ -MT in *B. glabrata* might have been an evolutionary process toward the need of more specificity for detoxification of non-essential metals (i.e., primarily Cd). This hypothesis is congruent with the evolutionary theory of multi-domain MTs in bivalves. In bivalves, development of Cd-preferring MT has been proposed to be based on the conversion of a Cu-preferring β -domain to the Cd-preferring α -domain by the acquisition of additional Cys, followed by subsequent domain duplications (Jenny et al. 2004; Jenny et al. 2006). On the other hand, in pulmonate gastropods, it has been widely proposed insofar that further domain duplication from the prototypic $\beta_2\beta_1$ -MT form has unlikely happened. Instead, specific MT isoforms with different metal selectiveness in pulmonate gastropods have been achieved mainly through the composition changes of non-Cys aa (Jenny et al. 2016; Palacios et al. 2011). However, from the new evidence in this study, domain duplication giving rise to large MTs should be considered as one of the important mechanisms permitting pulmonate MTs to achieve more specificity for their cognate heavy metals. Taking into account that $\beta_2\beta_2\beta_2\beta_{1P}$ -MT-originated β_2 -domains display much closer relationship among themselves than with the Cd-MT-originated β_2 -domain in the phylogenetic analysis, it is likely that the divergence to the $\beta_2\beta_2\beta_2\beta_{1P}$ -MT in the *B. glabrata* genome might have occurred through a separate path independent of the pathway for generating the Cd-MT (Fig. 2). Hence, exposure experiments to examine metal selectiveness or binding property of $\beta_2\beta_2\beta_2\beta_{1P}$ -MT domains would be helpful to test this hypothesis.

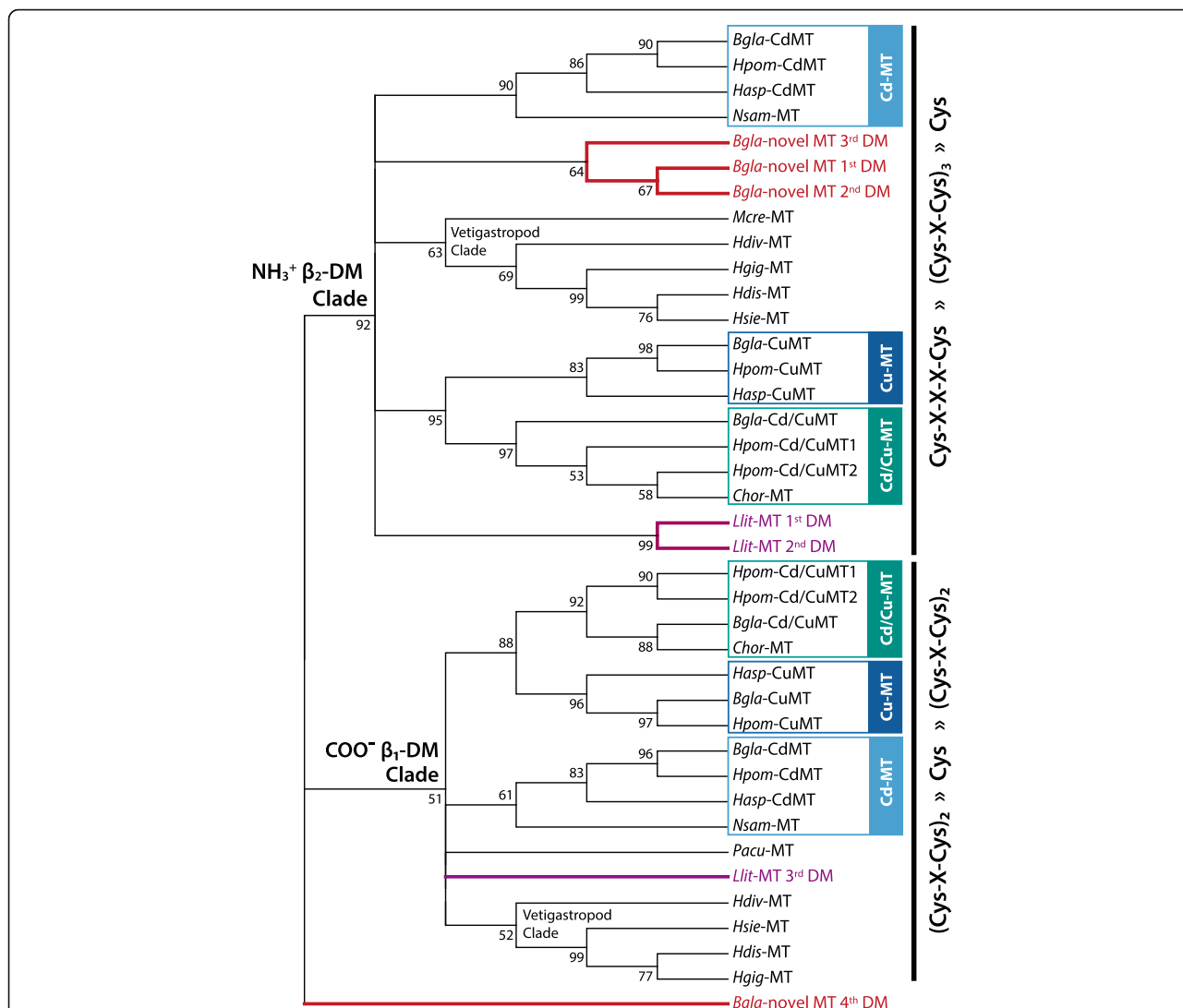


Fig. 1 Neighbor-joining tree showing the phylogenetic relationship among gastropoda MT β -domains (DM) analyzed with MEGA7 software (ver. 7.0.21) (Kumar et al. 2016). Based on the bootstrap tests (1000 replicates), only the clades supported by higher than 50% (condensed tree cutoff value = 50%) are visualized. For MT abbreviations and species, refer to Additional file 1: Table S1. In the NJ tree, two distinct clades, respectively, comprising of β_1 - and β_2 -domains indicate the presence of two ancestral β -domains in the gastropod lineage (Jenny et al. 2016). Typical arrangement patterns of Cys motifs for gastropod β_1 - and β_2 -domains are noted at the right side of the tree. Most gastropod MTs are structured as N-terminal β_2 -domain linked to C-terminal β_1 -domain. However, noticeably, some gastropod MTs are proposed to possess more than two β -domains (likely caused by tandem duplication) based on newly recognized evidences from *B. glabrata* MT ($\beta_2\beta_2\beta_2\beta_{1P}$ -like shape) and *L. littorea* MT ($\beta_2\beta_2\beta_1$ -structure). Evidence for the β_2 -domain duplication from the sequence alignment analysis can also be referred to in Additional file 1: Figure S2

The other evidence for the β_2 -domain duplication in the gastropod MTs is observable in the periwinkle (*L. littorea*; Caenogastropoda; Hypsogastropoda) MT. In the molecular phylogenetic tree, a subclade consisting of two closely affiliated *L. littorea* β -domains (first and second domains numbered from the N-terminal) was placed within the gastropod β_2 -clade, whereas the third β -domain of the *L. littorea* MT was positioned in the gastropod β_1 -clade (Fig. 1). Based on the phylogenetic separation between first, second, and third β -domains indicates that the *L. littorea* MT is comprised of two

successive β_2 -domains from the N-terminal that is linked to the C-terminal β_1 -domain (i.e., $\beta_2\beta_2\beta_1$ -MT). Like the *B. glabrata* $\beta_2\beta_2\beta_2\beta_{1P}$ -MT above, the first and second β_2 domains in the *L. littorea* MT share high sequence similarity including the conserved Cys motifs, suggesting that they might have evolved from a tail-to-head tandem duplication event (Additional file 1: Figure S2). Further efforts to exploit potential paralog isoforms from this species or closely related species are needed to hypothesize potential factor(s) to drive the domain duplication in *L. littorea* MT.

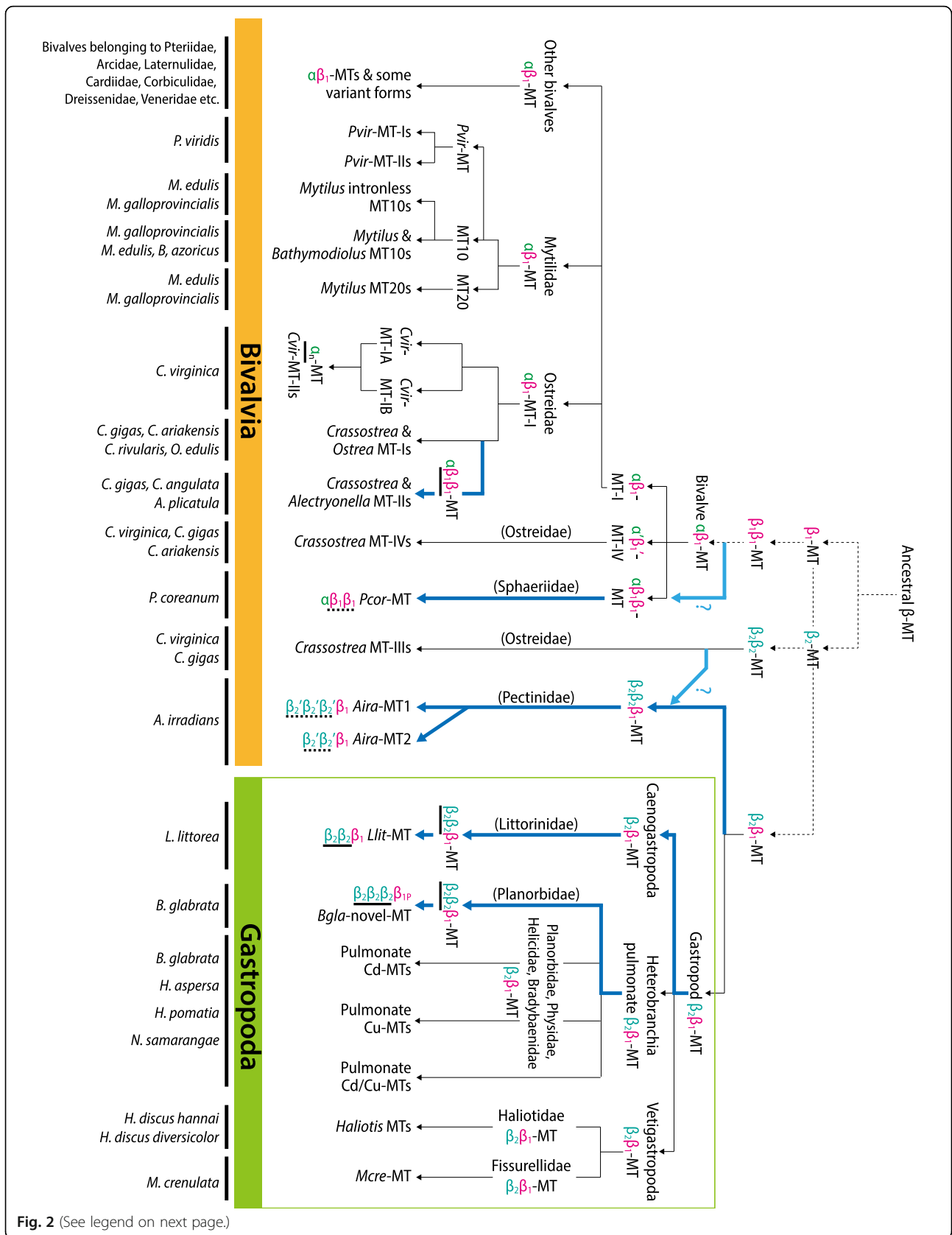


Fig. 2 (See legend on next page.)

(See figure on previous page.)

Fig. 2 A schematic representation to suggest the revised hypothesis including the newly proposed evolutionary paths (*thickened arrows*) for the divergence of MTs in the molluskan lineage. For MT abbreviations and species, refer to Additional file 1: Table S1. Ancestral β -domain-MT has been duplicated to early $\beta_1\beta_1$ - (eventually to $\alpha\beta_1$), $\beta_2\beta_2$ - and $\beta_2\beta_1$ -structured MTs and then further diverged into various shapes of extant MTs in the bivalve and gastropoda classes through domain duplication and/or aa substitutions in species/lineage-specific fashion. For MT isoforms with more than two metal-binding domains, the presumed duplicated domains are *underlined* with either a *solid line* (for homologous duplication giving rise to tandem array of domains with high sequence similarity) or *dashed line* (for heterologous duplication resulting in domains sharing little sequence similarity). Representative species examples for each MT type are noted *below* the schematic representation

Postmortem studies have claimed that most gastropod MTs would be very conservative in the Cys arrangements as a $\beta_2\beta_1$ -domain structure (Jenny et al. 2016; Berger et al. 1997; Dallinger et al. 1993). On the contrary, substitutions/replacements of non-Cys aa residues while retaining the $\beta_2\beta_1$ -frame have been thought as the major process for the evolutionary divergences of MTs in this primitive class Gastropoda. The Cu homeostatic requirements (thought to be mainly operated by β -domains) from the use of hemocyanin as a respiratory pigment in these gastropods, which is not present in oysters, has also been proposed as one of plausible factors responsible for the lack of divergently duplicated domains in gastropod MTs (Jenny et al. 2016; Berger et al. 1997; Perez-Rafael et al. 2012; Perez-Rafael et al. 1844). However, the present study claims that the formation of large MT with more than two domains should not be a bivalve-exclusive episode. It might have been an important path allowing some gastropod MTs to better modulate metal specificity in response to variations occurred in their habitat environments (Fig. 2). For both examples (*B. glabrata* MT and *L. littorea* MT), the target domain that has undergone duplication is the β_2 -domain. Hence, novel or specified functions (e.g., detoxification of non-essential metals) could be assigned to duplicated β_2 -domains while original and fundamental roles (e.g., Cu homeostatic regulation) are retained in the β_1 -domain. Extra metal-binding residues offered by duplicated β_2 -domains may also be potentially advantageous to strengthen further capacity of both metal reservation and resistance to excessive metal ions. Taken together, hypothesis for evolutionary mechanism to mold gastropod MTs should be revised taking into account the inclusion of this novel path featured by the duplication of β_2 -domain.

On the other hand, unlike in Heterobranchia and Caenogastropoda, the clear sign for taxa-specific domain multiplication has not been yet identified in Vetigastropoda species (abalone and limpet) (Lee and Nam 2016; Lieb 2003). Although the characterization of MT in this taxonomic group has been very limited, vetigastropod species have reportedly shown only a single MT isoform (i.e., $\beta_2\beta_1$ -MT) within a given species (Fig. 2). Currently, it is unclear if vetigastropods possess functionally or structurally diverged paralogs. However, a recent study has reported that the abalone (*Haliotis discus hannai*)

MT would be responsive to not only various heavy metals including Cu, Cd, and Zn but also non-metal stimulating stress treatments such as induced hypoxia, immune challenge and heat shock (Lee and Nam 2016; Guo et al. 2013). Based on this observation, the MT, at least in this vetigastropod species, is thought to have evolved to play readily multifunctional roles in diverse pathways involved in stress physiology.

Updates in ostreidae and mytilidae MTs

The evolutionary theory of MT based on domain duplication has been the most comprehensively highlighted in the *Crassostrea* species, particularly in *C. virginica*. Two ancestral β -domains (i.e., β_1 and β_2) appear to have diverged to produce two different structural MT isoforms, i.e., $\alpha\beta_1$ -MT and $\beta_2\beta_2$ -MT in the oysters belonging to Ostreidae (Jenny et al. 2004; Jenny et al. 2006). The ancestral β_1 -domain appears to have duplicated to produce a two-domain-structured MT that ultimately led to the evolution of the $\alpha\beta_1$ -structured MTs, which is observable in the *Crassostrea* species MT-Is and MT-IVs. On the other hand, the *Crassostrea* species MT-IIIs reveal the typical $\beta_2\beta_2$ -domain structure, which might have been a descendant shape resulted from the duplication event of a single ancestral β_2 -domain (Fig. 2). In the reconstructed phylogenetic tree, Ostreidae β -domain sequences are placed on one of two main clades (i.e., either β_1 - or β_2 -domain clade), although several subclades within the β_1 -clade are not supported by high confidence values (Additional file 1: Figure S7). *Crassostrea* MT-IVs are closely related each other and distinguished from MT-I/-II isoforms within the β_1 -clade, suggesting the early divergence between MT-IV and MT-I/-II families from the prototypic $\alpha\beta_1$ -MT form. Also within the β_1 -clade, it is notable that the *C. virginica* MT isoforms have a tendency to be separately clustered from MTs from other *Crassostrea* species (such as *C. gigas*, *C. ariakensis*, *C. rivularis*, and *C. angulata*) as seen in both MT-I and MT-IV groups. It may suggest the divergence of these MT families during speciation in the genus *Crassostrea* to make *C. virginica* to be distinguished from other *Crassostrea* species (Jenny et al. 2016) (Additional file 1: Figure S7 and Additional file 1: Figure S8). The most apparent difference between *C. virginica* and other *Crassostrea* species is found in the

MT-II family. *C. virginica* MT-IIIs have been reported to the most recently evolved isoforms, which have formed through the loss of β -domain, giving rise to the sole α -domain structured MTs (Additional file 1: Figure S8). However, this divergence pattern in *C. virginica* is clearly contrasted by the duplication of β_1 -domain in *C. gigas* MT-IIIs (i.e., $\alpha\beta_1\beta_1$ -structured MT), suggesting the different evolutionary paths of the MT-II between the two closely related species belonging to the same genus (Jenny et al. 2004; Tanguy and Moraga 2001). Such a solely α -domain-based structure has been found only in *C. virginica* insofar, whereas $\alpha\beta_1\beta_1$ -domain structure has also been observed similarly in other *Crassostrea* species and non-*Crassostrea* species (Additional file 1: Figure S7). Meanwhile, the β_2 -domain clade comprising of *C. virginica* and *C. gigas* MT-III isoforms shows a monophyletic topology, indicating the divergence from a common ancestral $\beta_2\beta_2$ -MT origin. However, within the β_2 -clade, two subclades are characterized by the first and second β_2 -domains rather than by species. All the N-terminally present β_2 -domains are clustered together in a subclade while all the C-terminally present β_2 -domains in other subclade (see also (Jenny et al. 2016)). This finding may be indicative of that the first and second β_2 -domains of the *Crassostrea* MT-IIIs might appear to have originated before the separation of the two oyster species (Fig. 2).

Mytilidae species represent uniformly the $\alpha\beta_1$ -domain structure (known as the prototypic shape of bivalve MT). Some mytilid species have been reported to possess the intronless, relatively short MTs in their genomes. The presence of intronless MT genes has been proposed as the organisms' strategic means for the efficient response to changes of cellular metal circumstances through the rapid transcription of MT genes (Leignel et al. 2005). Molecular phylogenetic analysis of Mytilidae MT domains generated trees consistently comprising of three main clades; clades for mytilid MT10s, *P. viridis* MTs and mytilid MT20s (Additional file 1: Figure S9). In congruent with the previous phylogenetic results using the entire MT polypeptide region, present phylogenetic analyses using separate domains also suggest the early divergence of *P. viridis* MTs from the other mytilid orthologs (Leung et al. 2014). In the Mytilidae lineage, a critical event before speciation is the divergence of MT10 and MT20 forms (Aceto et al. 2011) (Fig. 2). As compared to MT10, MT20s are characterized by the acquisition of additional Cys residues (i.e., a Cys-Cys doublet) in their α -domains (Leignel and Lallier 2006). It could be thought as a process to prepare the paralogue MT varieties with better execution in the detoxification of non-essential metals, since more Cys residues are generally taken into account for enhanced capability for sequestering the toxic metals (i.e., metal tolerance). This hypothesis could be supported by the fact that MT20 would be more preferentially associated or exclusive reacted with non-

essential metals than MT10 (Leung et al. 2014; Lemoine et al. 2000; Vergani et al. 2007). Conversely, in some environmental situations, the early $\alpha\beta_1$ -MT might have diverged into functionally differentiated isoforms in the Mytilidae: MT10 to execute primarily the homeostatic regulation of physiologically relevant metals and MT20 to function in the detoxification of trace metals (Fig. 2).

Novel paths for domain duplication in bivalve MTs

Exploitation of genetic determinants for MTs from other bivalve taxa has often showed the species (or lineage)-specific variations in MT structure. However, currently limited volume of knowledge on these MTs still hurdles to hypothesize the evolutionary mechanism of non-canonical MT forms in detail. Reconstruction of molecular phylogenetic trees in this study displays two main clades: one is a large clade comprising of β_1 -domains from various taxa and the other is a small clade consisting of five presumed β_2 -domain sequences deciphered from two *A. irradians* (Pectinidae) paralogue MTs (Fig. 3). Within a former β_1 -clade, paralogue isoforms from a given single species (e.g., *R. philippinarum* MT1/MT2 and *L. elliptica* MT10a/10b) formed subclades supported by high bootstrap values. Similarly, several subclades consisted of orthologs from closely related species belonging to the same genus ([e.g., MTs from genus *Meretrix* (Chang et al. 2007; Wang et al. 2010; Jiang et al. 2016) and genus *Cerastoderma* (Desclaux-Marchand et al. 2007; Ladhar-Chaabouni et al. 2009; Paul-Pont et al. 2012)). Collectively, it suggests that they might have evolved from recent divergence at species or genus levels. In contrast, some paralogue MT isoforms are distantly placed in the phylogenetic tree, although they are placed in the same β_1 -clade. Such a distant relationship is found in the genus *Hyriopsis* where MT1 and MT2 paralogs are not affiliated depending upon species. Although nomenclatures MT1 and MT2 are not established clearly in this two species, an isoform of *H. cumingii* MT (GQ184290) is closely related with *H. schlegelii* MT1 ortholog (KJ019820), rather than its paralogue isoform (FJ861993). This finding may indicate that the divergence between MT1 and MT2 might have occurred earlier than the speciation of the two *Hyriopsis* species (Yang et al. 2014; Wang et al. 2016).

From the present molecular phylogenetic analysis, novel paths of MT evolution through domain duplication giving rise to large-sized MTs with more than two metal-binding domains could be proposed (Fig. 3). Evidences come from two bivalve species: one is *P. coreanum* (Sphaeriidae) (Baek et al. 2009) and the other is *A. irradians* (Pectinidae) (Liu et al. 2006; Wang et al. 2009). In the *P. coreanum*, the two putative β -domains are placed in the β_1 -clade. Within the β_1 -clade, the second (numbered from the N-terminal) β_1 -domain of *P. coreanum* MT is found to form a

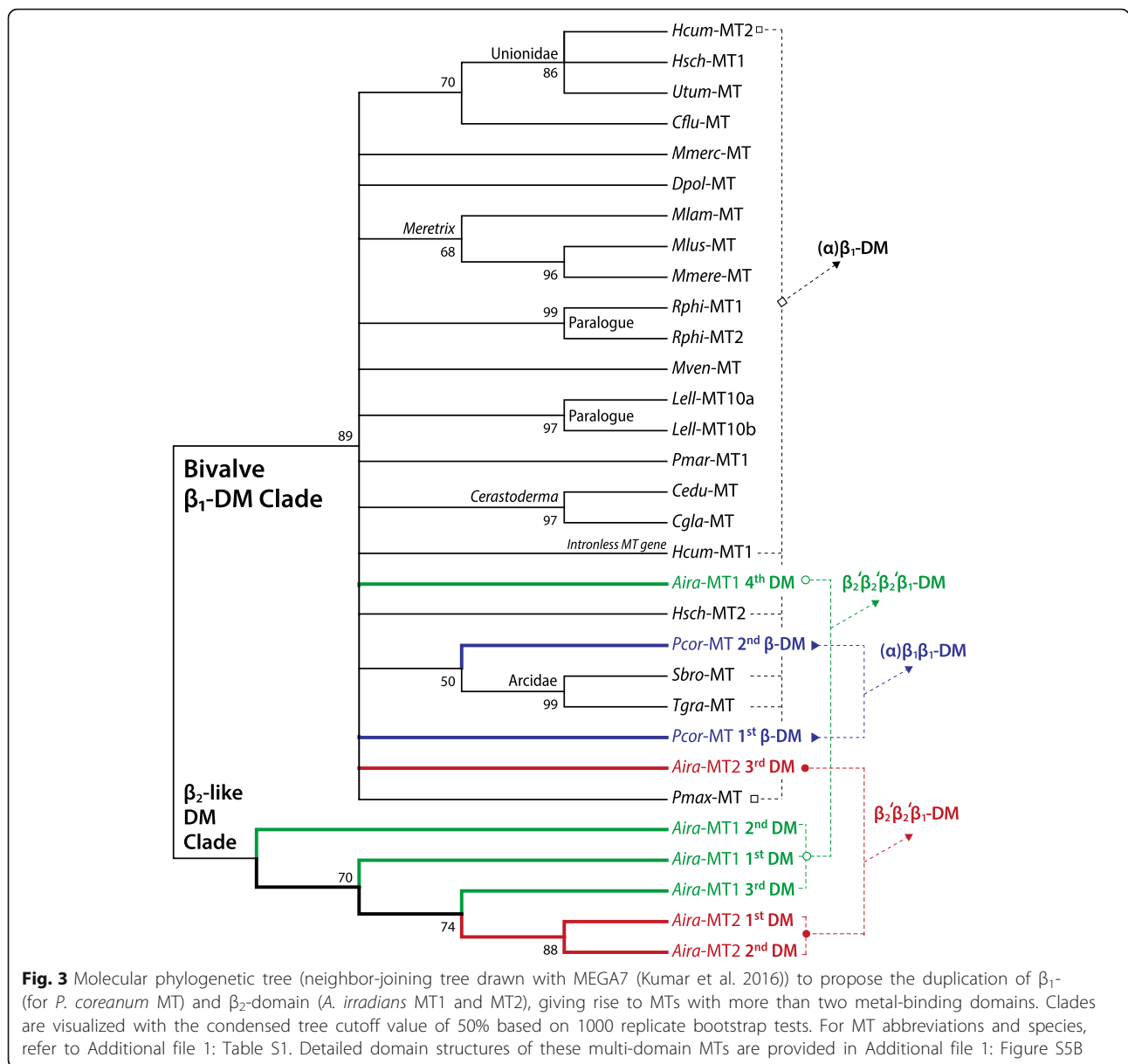


Fig. 3 Molecular phylogenetic tree (neighbor-joining tree drawn with MEGA7 (Kumar et al. 2016)) to propose the duplication of β_1 - (for *P. coreanum* MT) and β_2 -domain (*A. irradians* MT1 and MT2), giving rise to MTs with more than two metal-binding domains. Clades are visualized with the condensed tree cutoff value of 50% based on 1000 replicate bootstrap tests. For MT abbreviations and species, refer to Additional file 1: Table S1. Detailed domain structures of these multi-domain MTs are provided in Additional file 1: Figure S5B

subclade with two β_1 -domains from Arcidae species [*Scapharca broughtonii* (FJ154101) and *Tegillarca granosa* (AY568678)]. Considering the N-terminally present putative α -domain, this *P. coreanum* MT could be designated $\alpha\beta_1\beta_1$ -structure. In the phylum Mollusca, the $\alpha\beta_1\beta_1$ -domain structure (i.e., duplication of β_1 -domain from the anticipated prototypic $\alpha\beta_1$ -MT) has been previously reported to be the *Crassostrea*-specific event (Tanguy and Moraga 2001). However, we have already proposed above that this event has also been true for non-*Crassostrea* oyster (i.e., *A. plicatula*; Ostreidae). Further, the present *P. coreanum* (Sphaeriidae) MT could indicate that this duplication process would not be limited to the Ostreidae. However, the evolutionary scheme for the domain duplication may be different between the two families

Ostreidae and Sphaeriidae (Fig. 2). In Ostreidae, the β_1 -domain seems to have evolved from a relatively recent gene duplication, resulting in a tandem array of the two homologous β_1 -domains. On the contrary, the newly proposed *P. coreanum* $\alpha\beta_1\beta_1$ -MT shows no apparent sequence homology between the two β_1 -domains. Possibly, the multiplication of β_1 -domain in Sphaeriidae might have been an earlier divergent event. Currently, the evolutionary route for the acquisition of additional β_1 -domain in *P. coreanum* MT is open to hypothesize. One possible scenario is the duplication of β_1 -domain from the prototypic $\alpha\beta_1$ -MT, followed by further divergence in non-Cys residues. Alternatively, the other possibility is that the more ancestral $\beta_1\beta_1$ -MT (not yet reported in extant bivalve MTs) might have acquired additional α -domain through

the duplication to $\beta_1\beta_1\beta_1$ -MT followed by the conversion of one of β_1 -domains to α -domain (Fig. 2). Molecular phylogeny of bivalve α -domains also shows that *P. coreanum* α -domain is independently placed without being affiliated with any other bivalve α -domain ortholog in a subclade (Additional file 1: Figure S8). Hence, further efforts to exploit paralogue isoforms from this species (*P. coreanum*) and/or similarly structured orthologs from its closely related species should be needed to get a deeper insight into the mechanism responsible for the emergence of $\alpha\beta_1\beta_1$ -MT in Sphaeriidae.

From the same molecular phylogenetic tree, putative domains from the two *A. irradians* MT isoforms (Liu et al. 2006; Wang et al. 2009) are positioned in either the main β_1 -clade or a small clade consisting of only *A. irradians* MT domains (Fig. 3). Two C-terminal domains respectively from *A. irradians* MT1 and MT2 are placed in the main β_1 -clade. On the other hand, the small clade are exclusively comprised of three putative β_2 -like domains from the MT1 (first to third domains predicted from the N-terminal side) and two from MT2 (first and second domains). *A. irradians* MTs represent non-canonical shape that does not perfectly match the known typical β -domain structure. Nevertheless, if the Cys distribution pattern is fundamentally considered, the multiplied domains in *A. irradians* MTs could be classified as β_2 -like structure (possibly designated β_2' -domain). Hence, the overall domain structures of *A. irradians* MT1 and MT2 could be designated $\beta_2'\beta_2'\beta_2'\beta_1$ -MT and $\beta_2'\beta_2'\beta_1$ -MT, respectively. In bivalve class, the MT proteins possessing multiple β -domains without α -domain have been reported only in the *Crassostrea* $\beta_2\beta_2$ -MT-IIIIs, and this structure has been considered to have developed from the early duplication of the ancestral β_2 -MT. However, *A. irradians* MTs represent a C-terminal β_1 -domain as well as the multiple β_2' -domains (Fig. 2). Two plausible, but untested, hypotheses may be possible regarding the evolution of such unusual domain structure in *A. irradians* MTs. One is the acquisition of C-terminal β_1 -domain in the ancestrally duplicated $\beta_2\beta_2$ -MT (*i.e.*, giving rise to $\beta_2\beta_2\beta_1$ -MT) followed by further divergent process resulting in current shapes of *A. irradians* MT isoforms. The other, more plausible, hypothetical path is the multiplication of β_2 -domain from the prototypic $\beta_2\beta_1$ -MT structure that is seen in most gastropod MTs (Jenny et al. 2016; Palacios et al. 2011). As described above, the present study has already noted that the multiplication of β_2 -domain (*i.e.*, $\beta_{2n}\beta_1$ -structure) might have been one of divergence mechanisms in certain gastropod taxa (Fig. 2). However, unlike the tandem duplication of homologous β_2 -domain in gastropod MTs, *A. irradians* MTs represent little sequence similarity among β_2' -domains, suggesting that the multiplication of these β_2' -domains in each *A. irradians* MT isoform may not be the recent duplication

event. Although need to be further challenged, the ancestral $\beta_2\beta_1$ -MT in the phylum Mollusca perhaps might have gone through two separate paths (Fig. 2). Namely, one might have been a preservative path from the ancestral $\beta_2\beta_1$ -MT to produce the conserved $\beta_2\beta_1$ -structured MTs which seen in most extant gastropod MTs, although some exceptional gastropod species represent additional lineage-specific, recent duplication(s) of homologous β_2 -domain. On the other hand, the other path might involve earlier duplication of β_2 -domain from the ancestral $\beta_2\beta_1$ -MT, giving rise to multiple β_2 -like domains linked to the C-terminal β_1 -domain in certain bivalve taxa. Based on this, the extant shapes of *A. irradians* MT paralogs may reflect the consequences of differential rounds of β_2 -domain duplication from an ancestral $\beta_2\beta_1$ -MT. However, further validation is needed to test whether this divergent process might have occurred before or after speciation events in the Pectinoida lineage.

What are the functional or physiological implications of domain duplications (or multiplication) in molluscan MTs? The evolution of larger MT proteins has been proposed as a strategic means that might likely be advantageous for benthic organisms that are believed to experience a greater exposure to metals due to their ecological niche (Jenny et al. 2004; Jenny et al. 2006; Tanguy and Moraga 2001; Tschuschke et al. 2002). Although the paucity or limitation of the functional studies on such large MTs hurdles to hypothesize comprehensively the relevant mechanism(s) in detail, there have been some hypothetical evidences or suggestive assumptions. First, based on the heterologous expression assay by using the recombinant microbial systems, a couple of noteworthy experiments have shown that large-sized multi-domain-structured MT proteins would be able to confer greater Cd resistance of the hosts (Tanguy et al. 2001; Tschuschke et al. 2002). Second, several independent previous studies have claimed that amplification and/or tandem duplications of MT genes might have been an advantageous process to attain the strengthened ability of metal tolerance (Cho et al. 2009; Beach and Palmiter 1981; Maroni et al. 1987; Mehra et al. 1990; Stephan et al. 1994). Because the number of Cys residues in MT proteins has been taken into account as a fundamental factor to determine the number of metal ions bound or reserved by the MTs (Amiard et al. 2006), more Cys attained by domain duplications might be beneficial in the conference of metal resistance in a broad sense. Third, domain multiplication(s) accompanied with significant substitutions/replacements of non-Cys residues could offer a chance to confer some novel functions on large MTs. Because aa replacements on non-Cys residues in MT have been known to represent significant effects on metal-binding specificity and kinetic reactivity (Palacios et al. 2011; Pedersen et al. 1994; Kurasaki et al. 1997;

Munoz et al. 2000), such a divergent pattern (domain duplication together with significant aa substitutions) might also increase the kinds of metals reacted by these MT proteins. Taken together, the ancestral or prototypic MT protein has diverged into various isoforms with a great structural diversity in the phylum Mollusca. Structural diversifications driven by both domain duplication and aa replacements might have led certain subfunctionalization and/or neofunctionalization of MT proteins in an isoform-dependent fashion (Tanguy and Moraga 2001).

Conclusions

Phylum Mollusca represents a great structural diversity of MT, a core suite playing key roles in both homeostatic regulation of essential metals and detoxification of trace metals in living organisms. The structural diversity of molluscan MTs have been achieved essentially through the domain duplication events from an ancestral, singular domain-MT. Domain duplication have been followed by further diversification and selection toward needs for acquiring metal selectiveness, specialized novel function, and improved capacity of metal homeostasis/detoxification. With this viewpoint, novel paths for domain divergences of some gastropod and bivalve MT families proposed in this review could shed new light onto the revision and update of the hypothesis for evolutionary differentiation of MTs in the molluscan lineage.

Additional file

Additional file 1: Supplementary figures and tables. (PDF 433 kb)

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Authors' contributions

YKN designed this study, carried out bioinformatic analysis and drafted the manuscript. EJK carried out the management of sequence data, reference analyses, and polishing of the manuscript. Both authors read and approved the final manuscript.

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The authors declare that they have no competing interests.

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Ethics approval and consent to participate

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References

- Aceto S, Formisano G, Carella F, Vico GD, Gaudio L. The metallothionein genes of *Mytilus galloprovincialis*: genomic organization, tissue expression and evolution. *Mar Genom.* 2011;4:61–8.
- Amiard JC, Amiard-Triquet C, Barka S, Pellerin J, Rainbow PS. Metallothioneins in aquatic invertebrates: their role in metal detoxification and their use as biomarkers. *Aquat Toxicol.* 2006;76:160–202.
- Baek MK, Lee JS, Kang SW, Lee JB, Kang HJ, et al. Phylogenetic analysis based on metallothionein gene sequence of an indigenous species *Pisidium (Neopisidium) coreanum* in Korea. *Kor J Malacol.* 2009;25:153–60.
- Baršytė D, White KN, Lovejoy DA. Cloning and characterization of metallothionein cDNAs in the mussel *Mytilus edulis* L. digestive gland. *Comp Biochem Physiol C Pharmacol Toxicol Endocrinol.* 1999;122:287–96.
- Beach LR, Palmiter RD. Amplification of the metallothionein-I gene in cadmium-resistant mouse cells. *Proc Natl Acad Sci.* 1981;78:2110–4.
- Berger B, Dallinger R, Gehrig P, Hunziker PE. Primary structure of a copper-binding metallothionein from mantle tissue of the terrestrial gastropod *Helix pomatia* L. *Biochem J.* 1997;328:219–24.
- Binz PA, Kagi JHR. Metallothionein: molecular evolution and classification. In: Klaassen C, editor. *Metallothionein*. Basel: Birkhäuser; 1999. p. 7–13.
- Blindauer CA, Leszczyszyn OI. Metallothioneins: unparalleled diversity in structures and functions for metal ion homeostasis and more. *Nat Prod Rep.* 2010;27:720–41.
- Braun W, Wagner G, Worgotter E, Vasak M, Kagi JH, Wuthrich K. Polypeptide fold in the two metal clusters of metallothionein-2 by nuclear magnetic resonance in solution. *J Mol Biol.* 1986;187:125–9.
- Capasso C, Carginale V, Scudiero R, Crescenzi O, Spadaccini R, Temussi PA, Parisi E. Phylogenetic divergence of fish and mammalian metallothionein: relationships with structural diversification and organismal temperature. *J Mol Evol.* 2003;57:S250–7.
- Carpenè E, Andreani G, Isani G. Metallothionein functions and structural characteristics. *J Trace Elem Med Biol.* 2007;21:35–9.
- Chang YT, Jong KJ, Liao BK, Wu SM. Cloning and expression of metallothionein cDNA in the hard clam (*Meretrix lusoria*) upon cadmium exposure. *Aquaculture.* 2007;262:504–13.
- Chiaverini N, Ley MD. Protective effect of metallothionein on oxidative stress-induced DNA damage. *Free Radic Res.* 2010;44:605–13.
- Cho YS, Choi BN, Ha EM, Kim KH, Kim SK, Kim DS, Nam YK. Shark (*Scyliorhinus torazame*) metallothionein: cDNA cloning, genomic sequence, and expression analysis. *Mar Biotechnol.* 2005;7:350–62.
- Cho YS, Lee SY, Kim K-Y, Bang IC, Kim DS, Nam YK. Gene structure and expression of metallothionein during metal exposures in *Hemibarbus myloodon*. *Ecotoxicol Environ Saf.* 2008;71:125–37.
- Cho YS, Lee SY, Kim KY, Nam YK. Two metallothionein genes from mud loach *Misgurnus mizolepis* (Teleostei: Cypriniformes): gene structure, genomic organization, and mRNA expression analysis. *Comp Biochem Physiol B Biochem Mol Biol.* 2009;153:317–26.
- Cols N, Romero-Isart N, Bofill R, Capdevila M, Gonzalez-Duarte P, Gonzalez-Duarte R, Atrian S. In vivo copper- and cadmium-binding ability of mammalian metallothionein beta domain. *Protein Eng.* 1999;12:265–9.
- Cong M, Wu H, Liu X, Zhao J, Wang X, Lv J, Hou L. Effects of heavy metals on the expression of a zinc-inducible metallothionein-III gene and antioxidant enzyme activities in *Crassostrea gigas*. *Ecotoxicology.* 2012;21:1928–36.
- Dallinger R, Berger B, Hunziker PE, Birchler N, Hauer CR, Kägi JHR. Purification and primary structure of snail metallothionein. Similarity of the N-terminal sequence with histones H4 and H2A. *Eur J Biochem.* 1993;216:739–46.
- Desclaux-Marchand C, Paul-Pont I, Gonzalez P, Baudrimont M, Montaudouin X. Metallothionein gene identification and expression in the cockle (*Cerastoderma edule*) under parasitism (trematodes) and cadmium contaminations. *Aquat Living Resour.* 2007;20:43–9.
- Dondero F, Piacentini L, Banni M, Rebelo M, Burlando B, Viarengo A. Quantitative PCR analysis of two molluscan metallothionein genes unveils differential expression and regulation. *Gene.* 2005;345:259–70.
- English TE, Storey KB. Freezing and anoxia stresses induce expression of metallothionein in the foot muscle and hepatopancreas of the marine gastropod *Littorina littorea*. *J Exp Biol.* 2003;206:2517–24.
- Faller P. Neuronal growth-inhibitory factor (metallothionein-3): reactivity and structure of metal–thiolate clusters. *FEBS J.* 2010;277:2921–30.

- Fang Y, Yang H, Wang T, Liu B, Zhao H, Chen M. Metallothionein and superoxide dismutase responses to sublethal cadmium exposure in the clam *Macra veneriformis*. *Comp Biochem Physiol C Toxicol Pharmacol*. 2010;151:325–33.
- Fang Y, Yang H, Liu B, Zhang L. Transcriptional response of lysozyme, metallothionein, and superoxide dismutase to combined exposure to heavy metals and bacteria in *Macra veneriformis*. *Comp Biochem Physiol C Toxicol Pharmacol*. 2013;157:54–62.
- Geffard A, Geffard O, Amiard JC, His E, Amiard-Triquet C. Bioaccumulation of metals in sediment elutriates and their effects on growth, condition index, and metallothionein contents in oyster larvae. *Arch Environ Contam Toxicol*. 2007;53:57–65.
- Guo F, Tu R, Wang W-X. Different responses of abalone *Haliotis discus hannai* to waterborne and dietary-borne copper and zinc exposure. *Ecotoxicol Environ Saf*. 2013;91:10–7.
- Gupta SK, Singh J. Evaluation of mollusc as sensitive indicator of heavy metal pollution in aquatic system: a review. *IIOAB*. 2011;2:49–57.
- Inoue K-I, Takano H, Shimada A, Satoh M. Metallothionein as an anti-inflammatory mediator. *Mediat Inflamm*. 2009;101659.
- Isani G, Carpenè E. Metallothioneins, unconventional proteins from unconventional animals: a long journey from nematodes to mammals. *Biogeosciences*. 2014;4:435–57.
- Jenny MJ, Ringwood AH, Schey K, Warr GW, Chapman RW. Diversity of metallothioneins in the American oyster, *Crassostrea virginica*, revealed by transcriptomic and proteomic approaches. *Eur J Biochem*. 2004;271:1702–12.
- Jenny MJ, Warr GW, Ringwood AH, Baltzegar DA, Chapman RW. Regulation of metallothionein genes in the American oyster (*Crassostrea virginica*): ontogeny and differential expression in response to different stressors. *Gene*. 2006;379:156–65.
- Jenny MJ, Payton SL, Baltzegar DA, Lozier JD. Phylogenetic analysis of molluscan metallothioneins: evolutionary insight from *Crassostrea virginica*. *J Mol Evol*. 2016;83:110–25.
- Jiang LJ, Maret W, Vallee BL. The glutathione redox couple modulates zinc transfer from metallothionein to zinc-depleted sorbitol dehydrogenase. *Proc Natl Acad Sci*. 1998;95:3483–8.
- Jiang LJ, Vasak M, Vallee BL, Maret W. Zinc transfer potentials of the alpha- and beta-clusters of metallothionein are affected by domain interactions in the whole molecule. *Proc Natl Acad Sci*. 2000;97:2503–8.
- Jiang GP, Cheng XY, Teng SS, Chai XL, Lin XG, Liu GX, Xiao GQ. Cloning and expression of metallothionein gene in *Meretrix lamarckii*. *Acta Hydrobiol Sin*. 2016;40:914–20.
- Khoo HW, Patel KH. Metallothionein cDNA, promoter, and genomic sequences of the tropical green mussel. *Perna viridis* *J Exp Zool*. 1999;284:445–53.
- Kumar S, Stecher G, Tamura K. MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Mol Biol Evol*. 2016;33:1870–4.
- Kurasaki M, Yamaguchi R, Linde Arias R, Okabe M, Kojima Y. Significance of alpha-fragments of metallothionein in cadmium binding. *Prot Eng*. 1997;10:413–6.
- Ladhar-Chaabouni L, Mokdad-Gargouri R, Denis F, Hamza-Chaffai A. Cloning and characterization of cDNA probes for the analysis of metallothionein gene expression in the Mediterranean bivalves: *Ruditapes decussatus* and *Cerastoderma glaucum*. *Mol Biol Rep*. 2009;36:1007–14.
- Le TTY, Zimmermann S, Sures B. How does the metallothionein induction in bivalves meet the criteria for biomarkers of metal exposure? *Environ Pollut*. 2016;212:257–68.
- Lee SY, Nam YK. Transcriptional responses of metallothionein gene to different stress factors in Pacific abalone (*Haliotis discus hannai*). *Fish Shellfish Immunol*. 2016;58:530–41.
- Leignel V, Laulier M. Isolation and characterization of *Mytilus edulis* metallothionein genes. *Comp Biochem Physiol C Toxicol Pharmacol*. 2006;142:12–8.
- Leignel V, Hardivillier Y, Laulier M. Small metallothionein MT-10 genes in coastal and hydrothermal mussels. *Mar Biotechnol*. 2005;7:236–44.
- Lemoine S, Bigot Y, Sellos D, Cosson RP, Laulier M. Metallothionein isoforms in *Mytilus edulis* (Mollusca, Bivalvia): complementary DNA characterization and quantification of expression in different organs after exposure to cadmium, zinc, and copper. *Mar Biotechnol*. 2000;2:195–203.
- Leung PTY, Park TJ, Wang Y, Che CM, Leung KMY. Isoform-specific responses of metallothioneins in a marine pollution biomonitor, the green-lipped mussel *Perna viridis*, towards different stress stimulations. *Proteomics*. 2014;14:1796–807.
- Lieb B. A new metallothionein gene from the giant keyhole limpet *Megathura crenulata*. *Comp Biochem Physiol C Toxicol Pharmacol*. 2003;134:131–7.
- Liu WQ, Ni DJ, Song LS, Wu LT, Xu W, Kong XY. Cloning and characterization of a metallothionein gene in bay scallop *Argopecten irradians*. *Oceanol Limnol Sin*. 2006;37:444–9.
- Lü D, Luo KY, Pan BP, Gao H. Expression of metallothionein and thioredoxin gene in *Cyclina sinensis* exposed to cadmium. *Oceanol Limnol Sin*. 2012;43:47–51.
- Lynes MA, Hidalgo J, Manso Y, Devisscher L, Laukens D, Lawrence DA. Metallothionein and stress combine to affect multiple organ systems. *Cell Stress Chap*. 2014;19:605–11.
- Mao H, Wang DH, Yang WX. The involvement of metallothionein in the development of aquatic invertebrate. *Aquat Toxicol*. 2012;110–111:208–13.
- Marie V, Gonzalez P, Baudrimont M, Boutet I, Moraga D, Bourdineaud JP, Boudou A. Metallothionein gene expression and protein levels in triploid and diploid oysters *Crassostrea gigas* after exposure to cadmium and zinc. *Environ Toxicol Chem*. 2006;25:412–8.
- Maroni G, Wise J, Young JE, Otto E. Metallothionein gene duplications and metal tolerance in natural populations of *Drosophila melanogaster*. *Genetics*. 1987;117:739–44.
- Mehra RK, Garey JR, Winge DR. Selective and tandem amplification of a member of the metallothionein gene family in *Candida glabrata*. *J Biol Chem*. 1990;265:6369–75.
- Munoz A, Petering DH, Shaw CF. 3rd. Structure-reactivity among metallothionein three-metal domains: role of noncysteine amino acid residues in lobster metallothionein and human metallothionein-3. *Inorg Chem*. 2000;39:6114–23.
- Nielson KB, Winge DR. Order of metal binding in metallothionein. *J Biol Chem*. 1983;258:13063–9.
- Nielson KB, Winge DR. Preferential binding of copper to the beta domain of metallothionein. *J Biol Chem*. 1984;259:4941–6.
- Palacios O, Pagani A, Perez-Rafael S, Egg M, Hockner M, Brandstatter A, Capdevila M, Atrian S, Dallinger R. Shaping mechanisms of metal specificity in a family of metazoan metallothioneins: evolutionary differentiation of mollusc metallothioneins. *BMC Biol*. 2011;9:4.
- Paul-Pont I, Gonzalez P, Montero N, Montaudouin X, Baudrimont M. Cloning, characterization and gene expression of a metallothionein isoform in the edible cockle *Cerastoderma edule* after cadmium or mercury exposure. *Ecotoxicol Environ Saf*. 2012;75:119–26.
- Pedersen KL, Pedersen SN, Højrup P, Andersen JS, Roepstorff P, Knudsen J, Depledge MH. Purification and characterization of a cadmium-induced metallothionein from the shore crab *Carcinus maenas* (L.). *Biochem J*. 1994;297:609–14.
- Perez-Rafael S, Monteiro F, Dallinger R, Atrian S, Palacios O, Capdevila M. *Cantareus aspersus* metallothionein metal binding abilities: the unspecific CaCd/CuMT isoform provides hints about the metal preference determinants in metallothioneins. *Biochim Biophys Acta*. 1844;2014:1694–707.
- Perez-Rafael S, Mezger A, Lieb B, Dallinger R, Capdevila M, Palacios O, Atrian S. The metal binding abilities of *Megathura crenulata* metallothionein (McMT) in the frame of Gastropoda MTs. *J Inorg Biochem*. 2012;108:84–90.
- Raspor B, Dragun Z, Erk M, Ivankovic D, Pavicic J. Is the digestive gland of *Mytilus galloprovincialis* a tissue of choice for estimating cadmium exposure by means of metallothioneins? *Sci Total Environ*. 2004;333:99–108.
- Sarkar A, Ray D, Shrivastava AN, Sarker S. Molecular biomarkers: their significance and application in marine pollution monitoring. *Ecotoxicology*. 2006;15:333–40.
- Serén N, Glaberman S, Carretero MA, Chiari Y. Molecular evolution and functional divergence of the metallothionein gene family in vertebrates. *J Mol Evol*. 2014;78:217–33.
- Stephan W, Rodriguez VS, Zhou B, Parsch J. Molecular evolution of the metallothionein gene Mtn in the melanogaster species group: results from *Drosophila ananassae*. *Genetics*. 1994;138:135–43.
- Tang RS, Xia JH, Wang YM, Gong SY, Yu DH. Analysis on cloning and sequence characteristics of metallothionein cDNA in *Pinctada maxima*. *J Anhui Agricult Sci*. 2009;37:2888–90.
- Tanguy A, Moraga D. Cloning and characterization of a gene coding for a novel metallothionein in the Pacific oyster *Crassostrea gigas* (CgMT2): a case of adaptive response to metal-induced stress? *Gene*. 2001;273:123–30.
- Tanguy A, Mura C, Moraga D. Cloning of a metallothionein gene and characterization of two other cDNA sequences in the Pacific oyster *Crassostrea gigas* (CgMT1). *Aquat Toxicol*. 2001;55:35–47.
- Tanguy A, Boutet I, Riso R, Boudry P, Auffret M, Moraga D. Metallothionein genes in the European flat oyster *Ostrea edulis*: a potential ecological tool for environmental monitoring? *Mar Ecol Prog Ser*. 2003;257:87–97.
- Tschuschke S, Schmitt-Wrede HP, Greven H, Wunderlich F. Cadmium resistance conferred to yeast by a non-metallothionein-encoding gene of the earthworm *Enchytraeus*. *J Biol Chem*. 2002;277:5120–5.
- Vergani L, Grattarola M, Borghi C, Dondero F, Viarengo A. Fish and molluscan metallothioneins: a structural and functional comparison. *FEBS J*. 2005;272:6014–23.

- Vergani L, Grattarola M, Grasselli E, Dondero F, Viarengo A. Molecular characterization and function analysis of MT-10 and MT-20 metallothionein isoforms from *Mytilus galloprovincialis*. *Arch Biochem Biophys*. 2007;465:247–53.
- Wang H, Zhang Q, Cai B, Li H, Sze KH, Huang ZX, Wu HM, Sun H. Solution structure and dynamics of human metallothionein-3 (MT-3). *FEBS Lett*. 2006; 580:795–800.
- Wang L, Song L, Ni D, Zhang H, Liu W. Alteration of metallothionein mRNA in bay scallop *Argopecten irradians* under cadmium exposure and bacteria challenge. *Comp Biochem Physiol C Toxicol Pharmacol*. 2009;149:50–7.
- Wang Q, Wang X, Wang X, Yang H, Liu B. Analysis of metallothionein expression and antioxidant enzyme activities in *Meretrix meretrix* larvae under sublethal cadmium exposure. *Aquat Toxicol*. 2010;100:321–8.
- Wang W-C, Mao H, Ma D-D, Yang W-X. Characteristics, functions and applications of metallothionein in aquatic vertebrates. *Front Mar Sci*. 2014;1:34.
- Wang C, Sheng J, Hong Y, Peng K, Wang J, Wu D, Shi J, Hu B. Molecular characterization and expression of metallothionein from freshwater pearl mussel, *Hyriopsis schlegelii*. *Biosci Biotechnol Biochem*. 2016;80:1327–35.
- Xiong Y, Chen Y, Ru B. The expressed alpha domain of mouse metallothionein-I from *Escherichia coli* displays independent structure and function. *Biochem Mol Biol Int*. 1998;46:307–19.
- Yang S, Wei M, Yang X, Wang H, He L, Li C. A novel metallothionein gene from mussel, *Hyriopsis cumingii*: Identification and expression under lanthanum exposure. *J World Aquacult Soc*. 2014;45:454–60.
- Zhang G, Fang X, Guo X, Li L, Luo R, Xu F, Yang P, Zhang L, Wang X, Qi H, et al. The oyster genome reveals stress adaptation and complexity of shell formation. *Nature*. 2012;490:49–54.

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