

RESEARCH ARTICLE

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Alterations of growth performance, hematological parameters, and plasma constituents in the sablefish, *Anoplopoma fimbria* depending on ammonia concentrations

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Abstract

Juvenile *Anoplopoma fimbria* (mean length 16.8 ± 2.2 cm, and mean weight 72.8 ± 5.4 g) were exposed for 2 months with different levels of ammonia (0, 0.25, 0.50, 0.75, 1.00, and 1.25 mg/L). Growth performances such as daily length gain, daily weight gain, condition factor, and hepatosomatic index were significantly decreased by ammonia exposure. Hematological parameters such as red blood cell (RBC) count, hematocrit, and hemoglobin were also significantly decreased. In plasma inorganic components, calcium and magnesium were significantly decreased by ammonia exposure. In plasma organic components, there was no alteration in cholesterol and total protein. In enzyme plasma components, glutamic oxalate transaminase (GOT) and glutamic pyruvate transaminase (GPT) were significantly increased. The results of this study indicated that ammonia exposure can induce significant growth reduction and blood biochemistry alterations of *A. fimbria*.

Keywords: Sablefish, Ammonia, Growth performance, Hematological parameters, Plasma components

Background

In Korea, aquaculture is a major industry in food security dimension because it can supply high-quality protein to the public by stably breeding aquatic organisms. However, ammonia hypergenesis by high density breeding in aquaculture environment is a critical environmental toxic factor to induce death. Exposure to excessive ammonia in aquatic animals induces depolarization in neuron because increased NH_4^+ displaces K^+ , which results in cell death in central nervous system. Therefore, it induces convulsions, coma, and death by the cell death (Thangam et al. 2014). In addition, acute ammonia exposure induces gill ventilation increase, equilibrium loss, convulsions, ionic balance failure, and hyper-excitability in aquatic animals (Kim et al. 2015).

Growth factor in aquaculture is one of the most basic and critical parameters to assess toxic effects by harmful substances in aquaculture environment. In fish exposed to toxic substances, growth performance is generally decreased by energy transition from use for growth and development to use for tissue damage recovery (Wendelaar Bonga, 1997). In aquatic environment, excessive ammonia concentrations can be accumulated in body fluids in fish, which results in growth inhibition, tissue erosion and degeneration, immune suppression, and high mortality (Liang et al., 2015).

Hematological and biochemical parameters in fish can be a critical indicator to assess alterations in circulatory system by toxic substances in external environment (Vinodhini and Narayanan, 2009). Ammonia especially affects hematological parameters in fish by blocking oxygen transfer from gill to blood (Thangam et al. 2014).

Sablefish, *Anoplopoma fimbria* used in this study is recognized as a high value fish species around the globe.

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In liberalization trend of the world market, aquatic products are also involved in the trend. Therefore, development in aquaculture of a high value fish species is urgent. The purpose of this study was to assess toxic effects of *A. fimbria* exposed to ammonia a part of complete sablefish culture, and to build standard breeding guidelines of sablefish aquaculture.

Methods

Experimental fish and conditions

Juvenile sablefish were obtained from Troutlodge Inc. in USA. During the acclimation period, the fish were fed diet twice daily and maintained on a 24-h dark cycle and constant condition at all times (Table 1). After acclimatization, 72 fish (body length, 16.8 ± 2.2 cm; body weight, 72.8 ± 5.4 g) were randomly selected for the study. The acclimation period commenced once the final temperature had been sustained for 24 h and animals were feeding, while showing no sign of stress. Ammonia exposure took place in tanks containing six fish per treatment group. Ammonia chloride (NH_4Cl) (Sigma, St. Louis, MO, USA) solution was dissolved in the respective tanks. The ammonia concentrations in the tanks were 0, 0.25, 0.50, 0.75, 1.00, and 1.25 mg/L, and actual ammonia concentrations are demonstrated in Table 2. Diluted 100 mg/L ammonia chloride (NH_4Cl) in 20 L glass tank to make respective concentrations (50, 0.25 mg/L, 100 ml; 0.50 mg/L, 150 ml; 0.75 mg/L, 200 ml; 1.00 mg/L, 250 ml; 1.25 mg/L). After the exposure experiment, feed was given at a rate of 2% body weight daily (as two 1% meals per day). There was no water flow rate. The tank water was thoroughly exchanged once per 2 days and made the same concentration in the respective 500 L circular tank. At the end of each period (at 1 and 2 months), animals were anesthetized in buffered 3-aminobenzoic acid ethyl ester methanesulfonate (Sigma Chemical, St. Louis, MO).

Growth

The weight and length of sablefish was measured just before exposure, at 1 and 2 months. Daily length gain, daily weight gain, condition factor, and hepatosomatic index (HSI) were calculated by the following method.

Table 1 The chemical components of seawater and experimental condition used in the experiments

Item	Value
Temperature (°C)	13.0 ± 1.0
pH	8.2 ± 0.5
Salinity (‰)	33.5 ± 0.6
Dissolved oxygen (mg/L)	7.8 ± 0.5
Chemical oxygen demand (mg/L)	1.21 ± 0.14

Table 2 Analyzed waterborne ammonia concentration from each source

Waterborne ammonia concentration (mg/L)	0	0.25	0.50	0.75	1.00	1.25
Waterborne ammonia concentrations	0	0.25	0.50	0.75	1.00	1.25
Actual ammonia concentrations	0.02 ± 0.01	0.28 ± 0.07	0.57 ± 0.14	0.81 ± 0.26	1.04 ± 0.32	1.32 ± 0.27

$$\text{Daily growth gain} = (W_f - W_i) / \text{day}$$

$$(W_f = \text{Final length or weight}, W_i = \text{Initial length or weight})$$

$$\text{Condition factor (\%)} = (W/L^3) \times 100$$

$$(W = \text{weight(g)}, L = \text{length(cm)})$$

$$\text{HSI} = (\text{liver weight} / \text{total fish weight}) \times 100$$

Blood samples and hematological assay

Blood samples were collected within 35–40 s through the caudal vein of the fish in 1-ml disposable heparinized syringes at the end of 1 and 2 months. The blood samples were kept at 4 °C until the blood parameters were completely studied. The total red blood cell (RBC) count, hemoglobin (Hb), concentration, and hematocrit (Ht) value were determined immediately. Total RBC counts were counted using optical microscope with hemo-cytometer (Improved Neubauer, Germany) after diluted by Hendrick's diluting solution. The Hb concentration was determined using Cyan-methemoglobin technique (Asan Pharm. Co., Ltd.). The Ht value was determined by the microhematocrit centrifugation technique. The blood samples were centrifuged to separate plasma from blood samples at 3000 g for 5 min at 4 °C. The plasma samples were analyzed for inorganic substances, organic substances, and enzyme activity using clinical kit (Asan Pharm. Co., Ltd.). In inorganic substances assay, calcium and magnesium were analyzed by the o-cresolphthalein-complexon technique and xylydyl blue technique. In organic substances assay, cholesterol and total protein were analyzed by enzyme method and by biuret technique. In enzyme activity assay, glutamic oxalate transaminase (GOT) and glutamic pyruvate transaminase (GPT) were analyzed by Kind-king technique.

Statistical analysis

The experiment was conducted in exposure periods for 2 months and performed triplicate. Statistical analyses were performed using the SPSS/PC+ statistical package (SPSS Inc, Chicago, IL, USA). Significant differences between groups were identified using one-way ANOVA and Tukey's test for multiple comparisons. The significance level was set at $P < 0.05$.

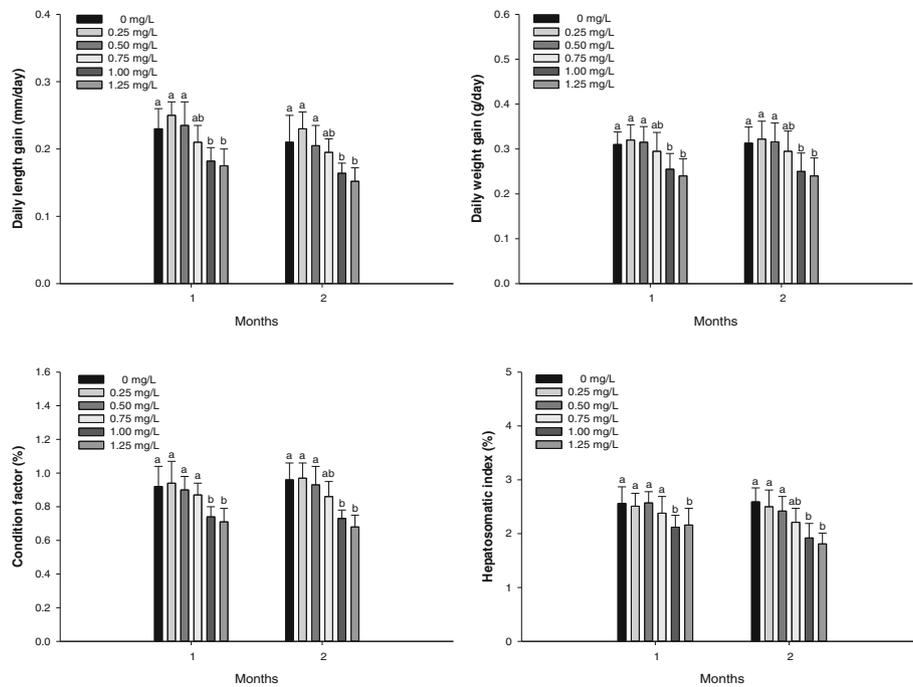


Fig. 1 Daily length gain, daily weight gain, condition factor, and hepatosomatic index of sablefish, *Anoplopoma fimbria* exposed to ammonia for 2 months. Vertical bar denotes a standard error. Values with different superscript are significantly different at 1 and 2 months ($P < 0.05$) as determined by Tukey's multiple range test

Results

Growth

No mortality was observed for the exposure periods. The growth performance, condition factor, and hepatosomatic index of *A. fimbria* is demonstrated in Fig. 1. Significant decreases in daily length gain and daily weight gain were observed at ammonia exposure greater than 1.00 mg/L both in 1 and 2 months. Condition factor was significantly decreased at ammonia exposure greater than 1.00 mg/L both in 1 and 2 months. Hepatosomatic index was also significantly decreased at ammonia exposure greater than 1.00 mg/L both in 1 and 2 months. However, there was no change in daily length, daily weight gain, and condition factor and

hepatosomatic index from 0 to 0.75 mg/L ammonia exposure after 1 and 2 months.

Hematological parameters

RBC count, hematocrit value, and hemoglobin concentration of *A. fimbria* exposed to different concentrations of waterborne ammonia are demonstrated in Fig. 2. RBC count was significantly decreased at ammonia exposure greater than 1.00 mg/L in 1 month and greater than 0.75 mg/L in 2 months. Hematocrit value was significantly decreased at ammonia exposure greater than 1.00 mg/L in 1 month and greater than 0.75 mg/L in 2 months. Hemoglobin concentration was significantly

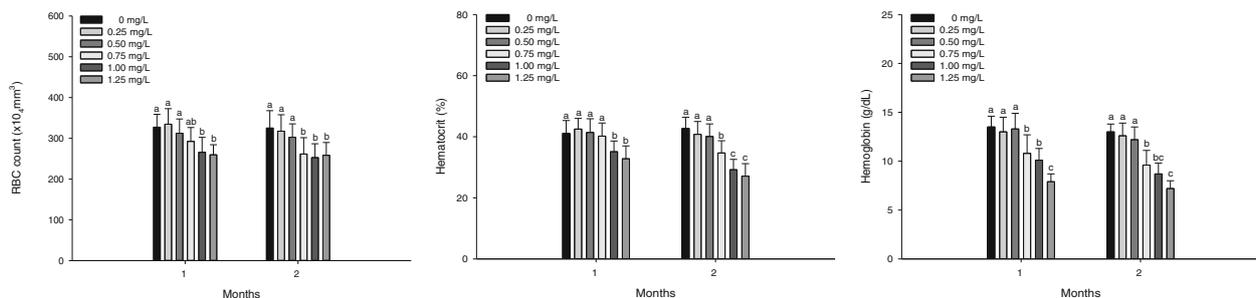


Fig. 2 RBC count, hematocrit, and hemoglobin of sablefish, *Anoplopoma fimbria* exposed to ammonia for 2 months. Vertical bar denotes a standard error. Values with different superscript are significantly different at 1 and 2 months ($P < 0.05$) as determined by Tukey's multiple range test

Table 3 Changes of inorganic plasma components in sablefish, *Anoplopoma fimbria* exposed to ammonia for 2 months

Parameters	Period (month)	Ammonia (mg/L)					
		0	0.25	0.50	0.75	1.00	1.25
Calcium (mg/dL)	1	1.84 ± 0.23 ^a	1.81 ± 0.26 ^a	1.75 ± 0.31 ^a	1.71 ± 0.30 ^{ab}	1.59 ± 0.25 ^b	1.55 ± 0.21 ^b
	2	1.81 ± 0.27 ^a	1.78 ± 0.23 ^a	1.62 ± 0.25 ^{ab}	1.64 ± 0.21 ^{ab}	1.49 ± 0.27 ^b	1.42 ± 0.24 ^b
Magnesium (mg/dL)	1	3.46 ± 0.42 ^a	3.51 ± 0.32 ^a	3.41 ± 0.27 ^a	3.29 ± 0.35 ^{ab}	2.89 ± 0.22 ^b	2.95 ± 0.32 ^b
	2	3.41 ± 0.36 ^a	3.46 ± 0.38 ^a	3.28 ± 0.41 ^{ab}	3.06 ± 0.25 ^b	2.74 ± 0.30 ^b	2.71 ± 0.24 ^b

Values are mean ± SE. Values with different superscript are significantly different at 1 and 2 months ($P < 0.05$) as determined by Tukey's multiple range test

decreased at ammonia exposure greater than 0.75 mg/L in 1 and 2 months.

Plasma components

Plasma inorganic components of *A. fimbria* are demonstrated in Table 3. Calcium was significantly decreased at ammonia exposure greater than 1.00 mg/L in 1 and 2 months. Magnesium was also significantly decreased at ammonia exposure greater than 1.00 mg/L in 1 month and greater than 0.75 in 2 months. Plasma organic components are demonstrated in Table 4. No alterations in cholesterol and total protein were observed by waterborne ammonia exposure. Plasma enzyme components are demonstrated in Table 5. GOT was significantly increased at ammonia exposure greater than 1.00 mg/L in 1 month and greater than 0.75 in 2 months. GPT was also significantly increased at ammonia exposure greater than 1.00 mg/L in 1 month and greater than 0.75 in 2 months.

Discussion

Ammonia exposure to fish is a critical environmental limited factor to inhibit growth performance by decreasing feed intake and feed utilization (Foss et al., 2003). Many authors reported that high concentrations of ammonia exposure induced growth inhibition of spotted wolfish, *Anarhichas minor* Olafsen (Foss et al., 2003), turbot, *Scophthalmus maximus* (Foss et al., 2009), Atlantic halibut, *Hippoglossus hippoglossus* (Paust et al., 2011). In this study, high concentrations of ammonia induced a significant decrease in growth of sablefish, *A. fimbria*, which may be due to energy transition from growth and development to detoxification. Hepatosomatic index (HSI) is considered as a critical indicator to evaluate

health status by toxic substance exposure (Datta et al., 2007), and HSI of sablefish, *A. fimbria* was significantly decreased by ammonia exposure.

Blood cells in fish are generated from hematopoietic tissues of kidney and spleen, and changes in hematological parameters indicate physiological effects by stress responses (Das et al., 2004). Jeney et al. (1992) suggest that high levels of ammonia exposure induce oxygen-free condition by increasing affinity of hemoglobin to combine with ammonia molecules, thereby elevating ammonia concentration in blood. Knoph and Thorud (1996) reported a significant decrease in RBC count and hematocrit of Atlantic salmon, *Salmo salar* exposed to ammonia. Das et al. (2004) also reported a significant decrease in hemoglobin of Mrigal carp, *Cirrhinus cirrhosus* exposed to ammonia. In this study, ammonia exposure caused a significant decrease in RBC count, hematocrit, and hemoglobin of sablefish, *A. fimbria*, which may be due to hematopoietic cell damage according to hypoxic status by ammonia exposure.

Calcium and magnesium in plasma inorganic components are critical indicators of osmotic pressure alterations, and these can be increased or decreased by environmental changes (Hur et al., 2001). Person-Le Ruyet et al., (2003) reported that ammonia exposure to turbot, *Scophthalmus maximus* induced changes in osmotic pressure by altering Na^+ , Cl^- , K^+ , Ca^{2+} concentrations in plasma. In this study, calcium and magnesium in sablefish, *A. fimbria* were significantly decreased by ammonia exposure, which indicate that ammonia exposure affected the osmotic ion regulation of sablefish. Cholesterol and total protein of plasma organic components in fish have been considered as a major component to assess fish health. However, there

Table 4 Changes of organic plasma components in sablefish, *Anoplopoma fimbria* exposed to ammonia for 2 months

Parameters	Period (month)	Ammonia (mg/L)					
		0	0.25	0.50	0.75	1.00	1.25
Cholesterol (mg/dL)	1	131.5 ± 20.5 ^a	139.1 ± 18.2 ^a	140.5 ± 21.3 ^a	137.2 ± 16.8 ^a	142.2 ± 21.1 ^a	146.2 ± 18.3 ^a
	2	135.1 ± 17.1 ^a	131.5 ± 17.1 ^a	138.2 ± 17.1 ^a	143.2 ± 21.3 ^a	148.3 ± 18.3 ^a	142.6 ± 20.2 ^a
Total protein (g/dL)	1	4.42 ± 0.53 ^a	4.35 ± 0.39 ^a	4.26 ± 0.45 ^a	4.51 ± 0.48 ^a	4.33 ± 0.37 ^a	4.59 ± 0.41 ^a
	2	4.36 ± 0.47 ^a	4.56 ± 0.35 ^a	4.37 ± 0.31 ^a	4.62 ± 0.52 ^a	4.24 ± 0.45 ^a	4.39 ± 0.51 ^a

Values are mean ± SE. Values with different superscript are significantly different at 1 and 2 months ($P < 0.05$) as determined by Tukey's multiple range test

Table 5 Changes of enzymatic plasma components in sablefish, *Anoplopoma fimbria* exposed to ammonia for 2 months

Parameters	Period (month)	Ammonia (mg/L)					
		0	0.25	0.50	0.75	1.00	1.25
GOT (karmen unit)	1	2.67 ± 0.33 ^a	2.72 ± 0.29 ^a	2.75 ± 0.35 ^a	2.81 ± 0.28 ^a	3.36 ± 0.41 ^b	3.45 ± 0.37 ^b
	2	2.72 ± 0.25 ^a	2.61 ± 0.32 ^a	2.91 ± 0.21 ^{ab}	3.24 ± 0.32 ^b	3.46 ± 0.40 ^{bc}	3.62 ± 0.41 ^c
GPT (karmen unit)	1	1.72 ± 0.24 ^a	1.79 ± 0.19 ^a	1.84 ± 0.18 ^a	1.93 ± 0.26 ^{ab}	2.19 ± 0.31 ^b	2.28 ± 0.27 ^b
	2	1.75 ± 0.28 ^a	1.82 ± 0.23 ^a	1.99 ± 0.28 ^a	2.23 ± 0.31 ^b	2.41 ± 0.35 ^c	2.40 ± 0.35 ^c

Values are mean ± SE. Values with different superscript are significantly different at 1 and 2 months ($P < 0.05$) as determined by Tukey's multiple range test

was no significant alteration in sablefish, *A. fimbria*. GOT and GPT in enzymatic plasma components can be easily increased by hepatic tissue damage, and these are used to evaluate hepatic tissue damage (Agrahari et al., 2007). Le Ruyet et al. (1998) reported that a significant increase in GOT and GPT of turbot, *Scophthalmus maximus* and seabream, *Sparus aurata* exposed to ammonia. In this study, GOT and GPT in sablefish, *A. fimbria* were significantly increased by ammonia exposure, which may be due to hepatic tissue damage by ammonia.

Conclusion

The results of this study indicate that ammonia exposure at the higher than proper concentrations affected growth performance and hematological parameters of sablefish, *A. fimbria*, and these changes should influence the health of sablefish, *A. fimbria*. In conclusion, ammonia concentrations at the higher than 0.75 mg/L can affect various physiological effects of sablefish, *A. fimbria*, and the high concentrations of ammonia exposure require special attention in sablefish aquaculture. In addition to this environmental study, various environmental standards should be established for stable sablefish aquaculture.

Abbreviations

GOT: Glutamic oxalate transaminase; GPT: Glutamic pyruvate transaminase; HIS: Hepatosomatic index; RBC: Red blood cell

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Authors' contributions

HJ, IK, and JM carried out the environmental toxicity studies and manuscript writing. JH, DH, CW, and JS participated in the design of the study and data analysis. JC participated in its design and coordination and helped to draft the manuscript. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Consent for publication

Not applicable

Ethics approval and consent to participate

All experimental animals used in this study were maintained under a protocol approved by the Institutional Animal Care and Use Committee of the Pukyong National University.

Disclosure

The dataset(s) supporting the conclusions of this article is not included in the article.

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