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Cloning and characterization of ADP-ribosylation factor 1b from the olive flounder *Paralichthys olivaceus*

So-Hee Son¹, Jin-Hyeon Jang¹, Hyeon-Kyeong Jo¹, Joon-Ki Chung² and Hyung-Ho Lee^{1*}

Abstract

Small GTPases are well known as one of the signal transduction factors of immune systems. The ADP-ribosylation factors (ARFs) can be classified into three groups based on the peptide sequence, protein molecular weight, gene structure, and phylogenetic analysis. ARF1 recruits coat proteins to the Golgi membranes when it is bound to GTP. The class I duplicated ARF gene was cloned and characterized from the olive flounder (*Paralichthys olivaceus*) for this study. PoARF1b contains the GTP-binding motif and the switch 1 and 2 regions. PoARF1b and PoARF1b mutants were transfected into a Hirame natural embryo cell to determine the distribution of its GDP/GTP-bound state; consequently, it was confirmed that PoARF1b associates with the Golgi body when it is in a GTP-binding form. The results of the qPCR-described PoARF1b were expressed for all of the *P. olivaceus* tissues. The authors plan to study the gene expression patterns of PoARF1b in terms of immunity challenges.

Keywords: ADP-ribosylation factor, Olive flounder, GTPase, Duplication, Immune system, Golgi complex

Background

The aquatic culture of the olive flounder (*Paralichthys olivaceus*) has been widespread in Korea. The farming of juvenile olive flounder, however, has caused a lot of problems due to the occurrence of various diseases (Ototake and Matsusato 1986; Park 2009). The juvenile flounder is difficult to manage and is weak against diseases, and the mortality rates have been economically damaging (Jee et al. 2001).

Small GTPases are well known as one of the signal transduction factors of immune systems (Narumiya 1996; Scheele et al. 2007). Some documents indicated that small GTPases are related to virus infection in the shrimp (Wu et al. 2007; Liu et al. 2009; Zhang et al. 2010). Also, the small GTPases of zebra fish have provided a firm basis of an innate immune system in vertebrates (Salas-Vidal et al. 2005). The authors therefore studied on ADP-ribosylation factor, which is a member of the GTP-binding proteins, from the olive flounder to

investigate the relation between cytoskeleton remodeling and the olive flounder immune system.

The ADP-ribosylation factor (ARF) proteins are small GTP-binding proteins, and they are involved in membrane dynamics and the regulation of actin cytoskeleton organization (D'Souza-Schorey and Chavrier 2006; Myers and Casanova 2008). The ARF can be classified into three groups based on the peptide sequence, protein molecular weight, gene structure, and phylogenetic analysis, as follows: class I including ARF1, ARF2, and ARF3; class II including ARF4 and ARF5; and class III including only ARF6 (Myers and Casanova 2008; Tsuchiya et al. 1991). The class I and class II ARFs are mainly associated with the Golgi complex, although they also function in endosomal compartments (Myers and Casanova 2008). In addition, ARF proteins were identified as activators of phospholipase D (PLD) (Luo et al. 1998). ARF1 was shown to recruit coat proteins to the Golgi membranes when it is bound to GTP (Balch et al. 1992). The hydrolysis and binding of GTP by ARF1 were originally linked to the assembly and disassembly of vesicle coats (Nie and Randazzo 2006).

The radiation of teleosts has been attributed to a genome-DNA event during the evolution of the teleosts

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(Venkatesh 2003). Although a number of ARFs, from those of micro-organisms to those of mammals, have been studied, a lack of studies on the duplicated ARF genes in the olive flounder persists. The authors therefore isolated and characterized one of the class I duplicated ARF genes.

Methods

cDNA cloning and phylogenetic analysis of *Paralichthys olivaceus* ARF1b

Total RNA was extracted by using GeneAll® Hybrid-R™ Total RNA (GeneAll Biotechnology Co., Ltd., Korea) following the manufacturer’s instructions from 12 tissues, including the brain, eye, gullet, heart, liver, stomach, muscle, kidney, spleen, pyloric caeca, intestine, and gill tissues, of healthy *Paralichthys olivaceus*. And then, we carried out 5’- and 3’-rapid amplification of cDNA ends (RACE) by using SMART™ RACE cDNA Amplification Kit (Clontech laboratories, Inc.) according to the manufacturer’s instructions. For obtaining full-length cDNA sequence, new gene-specific sense and antisense primers were designed (Table 1). The primers were used to PCR for obtaining full-length cDNA sequence. Nucleotide sequences and deduced amino acid sequence aligned with their respective homologues using Genetyx 7.0 software (GENETYX Corporation, Tokyo, Japan) and sequence alignment editor (BioEdit) (Hall 2011).

The phylogenetic tree was constructed using the Neighbor-Joining Method by MEGA6 (Tamura et al. 2013). Different DNA and protein sequences in Ensembl sequence database were used to carry out phylogenetic tree generation, sequence alignments, and database searching (Additional file 1) (Flicek et al. 2011).

Tissue distribution of PoARF1b by qPCR analysis

The tissue distribution of PoARF1b in different tissues was measured by RT-qPCR using a LightCycler 480 Real-Time PCR System (Roche, Mannheim, Germany) with LightCycler 480 SYBR green master I (Roche). The total RNA was extracted from the brain, gullet, eye, heart, stomach, liver, kidney, spleen, pyloric caeca, muscle, intestine, and gill from healthy *P. olivaceus* specimens. cDNA was synthesized with random hexamer primers and oligo(dT)18 using the PrimeScript™ 1st strand cDNA Synthesis Kit (TaKaRa), according to the manufacturer’s instructions. The specific primer for internal control was used 18s rRNA (Table 1) (Ahn et al. 2008). The quantitative real-time PCR followed program: pre-incubation at 95 °C for 5 min, 45 cycles at 95 °C for 10 s, 60 °C for 10 s, and 72 °C for 10 s. The qPCR reaction mixture consists of the following elements: 10 µl of 2× SYBR (Roche), 7.5 µl of SYBR water (Roche), 1 µl each of sense and antisense primers, and 0.5 µl diluted first-strand cDNA (diluted at 1:20). The ΔΔCt method

Table 1 Oligonucleotide primers used in PCR amplification of *ARF1b* of *P. olivaceus*; F, Forward; R, reverse

Primer name	5'-3' sequence	Information	
DgARF1b-F1	ATGGGDRMYWTBCKYWSC	Primers for cDNA library screening	
DgARF1b-F2	GACVACCATCYTGTACAARCTCAAAC		
DgARF1b-R1	CAYTTBVYDTTYTBAGCTKG		
DgARF1b-R2	CRTCYTCWGMCARCATTCCGCATC		
3’GSP-PoARF1b-F1	GTTGAGACAGTAGAGTACAGGAACATCAG	Primers for qPCR	
3’GSP-PoARF1b-F2	GGAGCGAATCGGTGAGGCGAGAGAGGAGC		
5’GSP-PoARF1b R1	CAGTCCCGCCGGGTGAGGTCGTGCAGGC		
5’GSP-PoARF1b R2	GCTCCTCTCTCGCCTCACCGATTGCTCC		
PoARF1b-RT-F	GCAGCAAAGTACTTCAAAGCCC		
PoARF1b-RT-R	CTCAGCCAGCATTCCGCATC		
Po18S rRNA-RT-For2	ATGGCCGTTCTTAGTTGGTG		GenBank accession no. EF126037.1
Po18S rRNA-RT-Rev2	CACACGCTGATCCAGTCAGT		
PoARF1b-ORF F	ATGGGACAGTCTTTAGCTGTTTAAAG		Primers for the construction of pEGFP Cl
PoARF1b-ORF R	TCATTTTATGTTCTTGTAGCTGGTTTAAAAG		
XhoI-PoARF1b-F	CCGCTCGAGCTATGGGACAGTCTTTA		
Xpnl-PoARF1b-R	CGGGGTACCTCATTATGTTCTTGTAGCTG	Primers for the site-directed mutation	
PoARF1bT30N-F	CTGCATGCTGGAAAGAACCACCATCTGTACAAA		
PoARF1bT30N-R	TTTGTACAGGATGGTGTCTTTCCAGCATGCAG		
PoARF1bQ70L-F	GTCGGTGGTCTGGACAAGATCAGGCCACTC		
PoARF1bQ70L-R	GAGTGGCCTGATCTTGTCCAGACCACCGAC		

was applied to calculate the data, and $2^{-\Delta\Delta Ct}$ method was applied to calculate the relative quantitative value (Giulietti et al. 2001).

Statistics

All qPCR data were statistically analyzed using SPSS 21 program (SPSS, Chicago, IL, USA). One-way ANOVA was used to study the PoARF1b expression, followed by Duncan's Multiple Range test. A p value with $p < 0.05$ was considered to be significant (Sokal and Rohlf 1969).

Cell culture and transfection

Hirame natural embryo cell line (HINAE) was grown in Leibovitz's L-15 medium (Gibco BRL, Grand Island, NY) containing 10% fetal bovine serum (Gibco) and 1% antibiotics (Gibco) at 20 °C (Kasai and Yoshimizu 2001). The transfection was performed using PolyPlus (JetPrime, New York, NY, USA) kit to transient transfection of PoARF1b and its mutants according to the manufacturer's instructions in six-well test plates. The PoARF1b and mutants were observed by EGFP fluorescence signal under confocal microscopy after 48 h post-transfection.

Site mutation of PoARF1b

PoARF1b(T30N) and PoARF1b(Q70L) were performed using QuikChange II Site-Directed Mutagenesis Kit (Agilent Technologies), according to the manufacturer's instructions (Wang and Malcolm 1999). For PoARF1b mutants, we used specific primers (Table 1). The pEGFP-C1 (Clontech) was used to construct the green fluorescent protein-fused PoARF1b and PoARF1b mutants.

The Golgi body in HINAE was stained using GOLGI ID® Green assay kit containing a Golgi apparatus-selective dye.

Results and Discussion

Cloning and sequence analysis of PoARF1b

To identify the initial sequence of PoARF1b, we obtained databases of other ARF1b by using Ensembl sequence data. These sequences were used to design the forward and reverse primers (Table 1). The initial sequence was obtained from PCR amplification of olive flounder cDNA including the brain, eye, gullet, heart, liver, muscle, stomach, kidney, spleen, pyloric caeca, intestine, and gill. The partial sequence was used for isolation of full-length flounder ARF1b by using 3' and 5' GeneRace with flounder ARF1b specific primers (Table 1). As the result, the full nucleotide sequence of PoARF1b is 1677 bp (GenBank accession no. KX668134).

The sequence comprised a 108 bp 5'-untranslated region (5'-UTR), a 544 bp coding region, and a 1025 bp 3'-untranslated region (3'-UTR). Also, the PoARF1b has

180 amino acid residues and the molecular weight is approximately 20,561 Da (Fig. 1a). PoARF1b contains GTP-binding motif, switch 1 and 2 regions (Fig. 1a) (Pasqualato et al. 2002). The GTP-binding motif is shaded with gray box and conserved sequence in other ARFs. The switch 1 and 2 regions were indicated with blue and red letters. The switch regions were significantly assumed to conformational change classical structural GDP/GTP switch that bind tightly to GTP but poorly or not at all to the GDP nucleotide (Pasqualato et al. 2002).

The aligned amino acid sequence appeared in PoARF1b had well conserved domains such as GTP-binding motif, switch 1 and 2 regions, and shared the high homology with ARFs from other species (Fig. 1b). It showed 90% homology with ARF1b from *Takifugu rubripes*.

Phylogenetic tree of PoARF1b

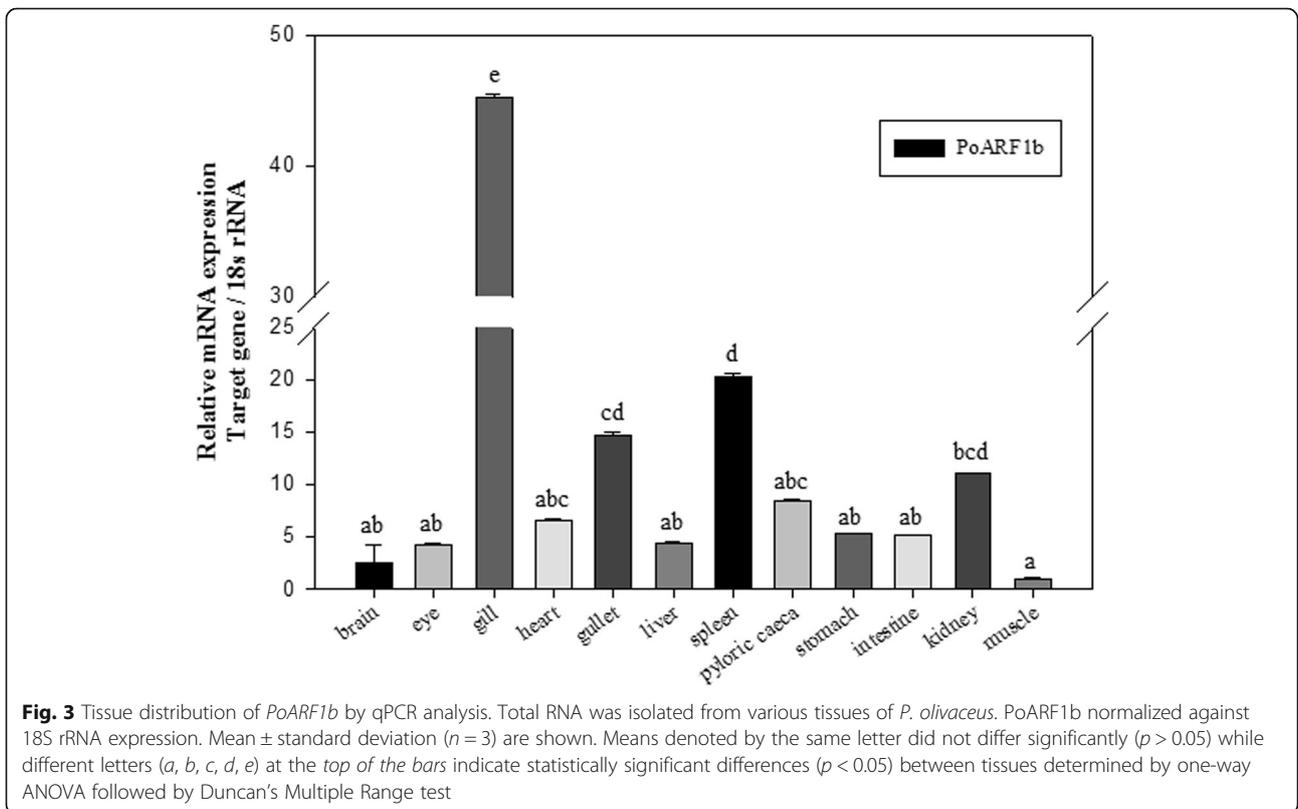
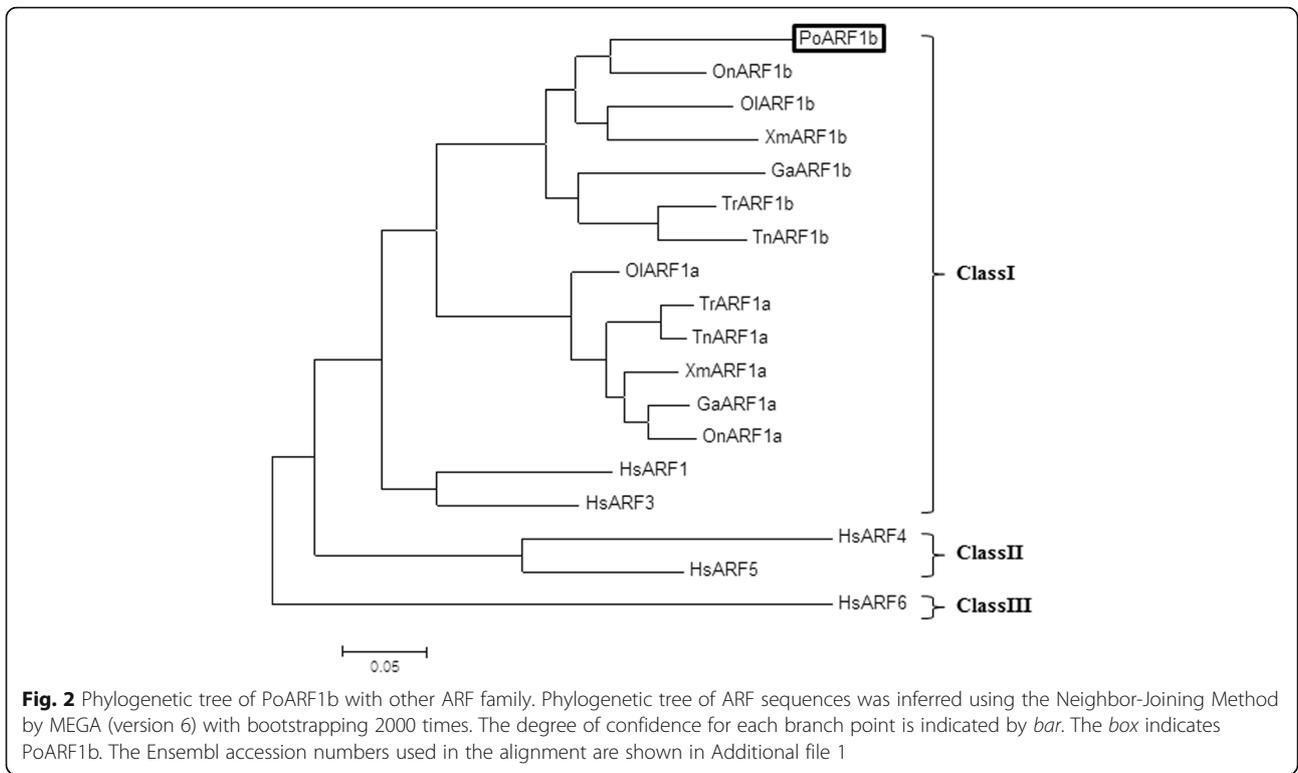
To determine the evolutionary relationship of PoARF1b with other ARFs, phylogenetic tree was performed using the Ensembl sequence data using the Neighbor-Joining Method by MEGA (version 6) with bootstrapping 2000 times (Flicek et al. 2011; Tamura et al. 2013). The result of phylogenetic tree was contained fish which was grouped with the tetrapod and human. The ARF tree consists of three major groups: (i) class I, (ii) class II, and (iii) class III. This result indicated that PoARF1b is closely related to ARF1b of class I (Fig. 2).

Tissue distribution of PoARF1b by qPCR analysis

Real-time PCR showed tissue distribution of PoARF1b. The results of qPCR-described PoARF1b was expressed for all mRNA transcripts in different organs which include the brain, gullet, eye, heart, stomach, liver, kidney, spleen, pyloric caeca, muscle, intestine, and gill (Fig. 3). The expression of the PoARF1b gene was the highest level in the gill and the lowest level in the muscle.

Site mutation analysis of PoARF1b

To determine the distribution of PoARF1b, PoARF1b, and PoARF1b mutants, those were constructed into pEGFP-C1 (Clontech) and transfected into HINAE cell. The punctuate morphology of PoARF1b-EGFP resembled the Golgi complex distribution in HINAE cells (Fig. 4b). This result examines PoARF1b that may act in the Golgi body like as human ARF1 complexed with GDP (Amor et al. 1994). Also, PoARF1b mutants examine the distribution which depends on each GDP or GTP-binding form. The mutants designed PoARF1b(T30N) and PoARF1b(Q70L) (Fig. 4a), according to other reports (Chavrier and Goud 1999; Teal et al. 1994). PoARF1b(T30N) was designed by exchanging the Thr amino acid at position 30 with Asn amino acid. It



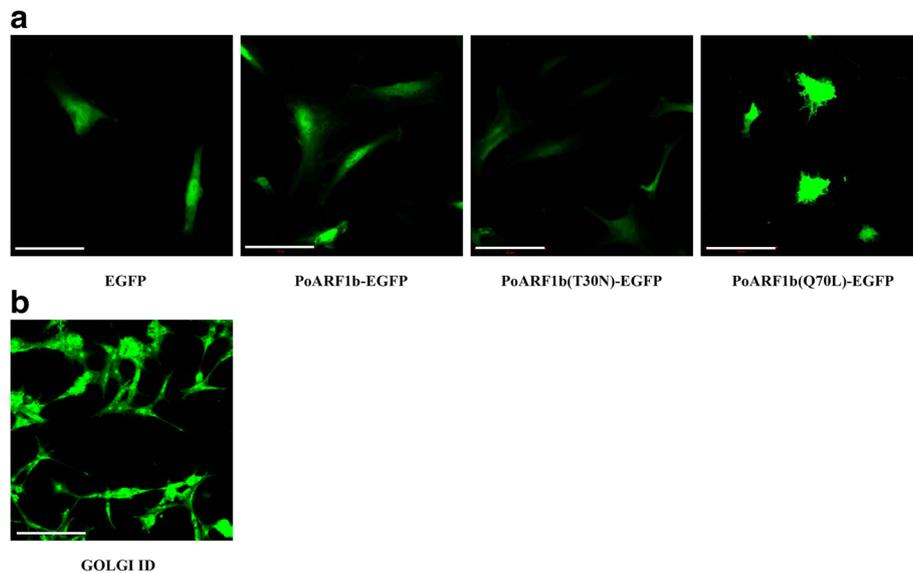


Fig. 4 a Punctuate morphology of PoARF1b and its mutants. The amounts of plasmids (2 μ g) were transfected into HINA E cells in 6 wells. EGFP, control; PoARF1b-EGFP, wild-type; PoARF1b(T30N)-EGFP, negative mutant; and PoARF1b(Q70L)-EGFP, positive mutant. **b** Intracellular distribution of the Golgi complex in HINA E cells using GOLGI ID. Bar means 50 μ m

identities from the other ARF isozymes (data not shown). PoARF shares approximately 85% with the identity of *Oreochromis niloticus* ARF1b (OnARF1b) and approximately 79% with the identity of *Gasterosteus aculeatus* ARF1b (GaARF1b). Also, PoARF shares 76% with the identity of *Homo sapiens* ARF1 (HsARF1). In addition, the phylogenetic tree showed that PoARF is more closely related to ARF1b than ARF1a. These results indicate that PoARF is PoARF1b. OnARF1b, which shares a high percentage with the identity of PoARF1b, shares 76% with the identity of HsARF1.

As is known, the PoARF1b is expressed in all of the tissues of the olive flounder. The PoARF1b mRNA has a high expression level in the gill and a low expression level in the muscle. This finding resembles the ARF1 expression from the shrimp (*Marsupenaeus japonicus*), which shows the lowest expression level in the muscle (Ma et al. 2010). It will be necessary to further study why such an outcome.

The Golgi-binding distribution of PoARF1b depends on the GTP- or GDP-bound state. The PoARF1b-EGFP showed a punctuate morphology that resembles the morphology of the Golgi body in HINA E cells (Fig. 4). The GOLGI ID of the Golgi bodies can be detected using a microscopy.

PoARF1b(T30N) showed a clearly disassembled punctuate morphology, and PoARF1b(Q70L) showed a more expanded morphology; these results resemble those of the mammalian ARF1. The results of this study indicate that PoARF1b functions within the Golgi complex.

Further studies are needed to be carried out to explain the highest expression of PoARF1b in gill.

Additional file

Additional file 1: Sequences used in sequence and phylogenetic analysis of PoARF1b. (TIF 493 kb)

Abbreviations

ARF: ADP-ribosylation factor; HINA E: Hirame natural embryo cell line; RACE: Rapid amplification of cDNA ends; UTR: Untranslated region

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Availability of data and materials

The dataset(s) supporting the conclusions of this article is(are) included within the article.

Authors' contributions

S-HS carried out the whole process, participated in the whole experiments and drafted the manuscript. J-HJ carried out the cDNA cloning and phylogenetic analysis. J-KC participated in its statistical analysis. HHL participated in the design of the study and coordination. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Consent for publication

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Ethics approval and consent to participate

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References

- Ahn SJ, Kim NY, Jeon SJ, Sung JH, Je JE, Seo JS, ... and Lee HH. Molecular cloning, tissue distribution and enzymatic characterization of cathepsin X from olive flounder (*Paralichthys olivaceus*). *Comp Biochem Physiol Part B Biochem Mol Biol*. 2008;151(2):203–212.
- Amor JC, Harrison DH, Kahn RA, Ringe D. Structure of the human ADP-ribosylation factor 1 complexed with GDP. *Nature*. 1994;372(6507):704–8.
- Balch WE, Kahn RA, Schwaninger R. ADP-ribosylation factor is required for vesicular trafficking between the endoplasmic reticulum and the cis-Golgi compartment. *J Biol Chem*. 1992;267(18):13053–61.
- Chavrier P, Goud B. The role of ARF and Rab GTPases in membrane transport. *Curr Opin Cell Biol*. 1999;11(4):466–75.
- D'Souza-Schorey C, Chavrier P. ARF proteins: roles in membrane traffic and beyond. *Nat Rev Mol Cell Biol*. 2006;7(5):347–58.
- Fliccek P, Amode MR, Barrell D, Beal K, Brent S, Carvalho-Silva D, ... and Gil L. Ensembl 2012. *Nucleic Acids Res*. 2011;gkr991.
- Giulietti A, Overbergh L, Valckx D, Decallonne B, Bouillon R, Mathieu C. An overview of real-time quantitative PCR: applications to quantify cytokine gene expression. *Methods*. 2001;25(4):386–401.
- Hall T. BioEdit: an important software for molecular biology. *GERF Bull Biosci*. 2011;2(1):6.
- Jee BY, Kim YC, Park MS. Morphology and biology of parasite responsible for scuticociliatosis of cultured olive flounder *Paralichthys olivaceus*. *Dis Aquat Org*. 2001;47(1):49–55.
- Johnson DS, Chen YH. Ras family of small GTPases in immunity and inflammation. *Curr Opin Pharmacol*. 2012;12(4):458–63.
- Kasai H, and Yoshimizu M. Establishment of two Japanese flounder [*Paralichthys olivaceus*] embryo cell lines. *Bulletin of Fisheries Sciences, Hokkaido University (Japan)*. 2001.
- Liu W, Han F, Zhang X. Ran GTPase regulates hemocytic phagocytosis of shrimp by interaction with myosin. *J Proteome Res*. 2009;8(3):1198–206.
- Luo JQ, Liu X, Frankel P, Rotunda T, Ramos M, Flom J, ... and Foster DA. Functional association between Arf and RalA in active phospholipase D complex. *Proc Natl Acad Sci*. 1998;95(7):3632–3637.
- Ma J, Zhang M, Ruan L, Shi H, Xu X. Characterization of two novel ADP ribosylation factors from the shrimp *Marsupenaes japonicus*. *Fish Shellfish Immunol*. 2010;29(6):956–62.
- Myers KR, Casanova JE. Regulation of actin cytoskeleton dynamics by Arf-family GTPases. *Trends Cell Biol*. 2008;18(4):184–92.
- Narumiya S. The small GTPase Rho: cellular functions and signal transduction. *J Biochem*. 1996;120(2):215–28.
- Nie Z, Randazzo PA. Arf GAPs and membrane traffic. *J Cell Sci*. 2006;119(7):1203–11.
- Ototake M, Matsusato T. Notes on Scuticociliata infection of cultured juvenile flounder *Paralichthys olivaceus*. *Bull. Natl. Res. Inst. Aquaculture*. 1986;9:65–68 (in Japanese).
- Park SI. Disease control in Korean aquaculture. *Fish Pathol*. 2009;44(1):19–23.
- Pasqualato S, Renault L, Cherfils J. Arf, Arl, Arp and Sar proteins: a family of GTP-binding proteins with a structural device for 'front-back' communication. *EMBO Rep*. 2002;3(11):1035–41.
- Radhakrishna H, Donaldson JG. ADP-ribosylation factor 6 regulates a novel plasma membrane recycling pathway. *J Cell Biol*. 1997;139(1):49–61.
- Salas-Vidal E, Meijer AH, Cheng X, Spaink HP. Genomic annotation and expression analysis of the zebrafish Rho small GTPase family during development and bacterial infection. *Genomics*. 2005;86(1):25–37.
- Scheele JS, Marks RE, Boss GR. Signaling by small GTPases in the immune system. *Immunol Rev*. 2007;218(1):92–101.
- Sokal RR, Rohlf FJ. The principles and practice of statistics in biological research. San Francisco: WH Freeman and company; 1969. p. 399–400.
- Tamura K, Stecher G, Peterson D, Filipski A, Kumar S. MEGA6: molecular evolutionary genetics analysis version 6.0. *Mol Biol Evol*. 2013;30(12):2725–9.
- Teal SB, Hsu VW, Peters PJ, Klausner RD, Donaldson JG. An activating mutation in ARF1 stabilizes coatamer binding to Golgi membranes. *J Biol Chem*. 1994;269(5):3135–8.
- Tsuchiya M, Price SR, Tsai SC, Moss J, Vaughan M. Molecular identification of ADP-ribosylation factor mRNAs and their expression in mammalian cells. *J Biol Chem*. 1991;266(5):2772–7.
- Venkatesh B. Evolution and diversity of fish genomes. *Curr Opin Genet Dev*. 2003;13(6):588–92.
- Wang W, Malcolm BA. Two-stage PCR protocol allowing introduction of multiple mutations, deletions and insertions using QuikChange Site-Directed Mutagenesis. *Biotechniques*. 1999;26(4):680–2.
- Wu W, Zong R, Xu J, Zhang X. Antiviral phagocytosis is regulated by a novel Rab-dependent complex in shrimp *Penaeus japonicus*. *J Proteome Res*. 2007;7(01):424–31.
- Zhang M, Ma J, Lei K, Xu X. Molecular cloning and characterization of a class II ADP ribosylation factor from the shrimp *Marsupenaes japonicus*. *Fish Shellfish Immunol*. 2010;28(1):128–33.

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